Glucose

	Livestock			
1	Identification of Petitioned Substance			
2				
3	Chemical Names:	18	Dextrose Solution 50%	
4	D-Glucose	19	Contained in trade name products such as:	
5	D-Glucopyranose	20	Hydra-Lyte Electrolyte Replacement	
6	D-Glc	21	Vitamins & Electrolytes "Plus" Oral Cal MPK	
7	D-Glucopyranoside	22		
8	Glc		CAS Numbers:	
9			50-99-7 (D-(+)-Glucose)	
10	Other Name:		2280-44-6 (D-Glucose)	
11	Glucose		54-17-1 (D-Glucose)	
12	Dextrose		77938-63-7 (D-Glucose monohydrate)	
13	Corn sugar			
14	Grape sugar		Other Codes:	
15			EINECS: 200-075-1	
16	Trade Names:		FDA UNII: 5SL0G7R0OK	
17	Dextrose 50%			
23				
24		Summary of Pet	itioned Use	

Glucose was included in the original National Organic Program (NOP) Final Rule in December 2000 (NOP, 2000).
Glucose is currently listed within the United States Department of Agriculture (USDA) organic regulations at 7
CFR §205.603(a)(13) as a synthetic substance allowed for use as a medical treatment in organic livestock
production. This technical report focuses on uses for glucose in organic livestock production, primarily to treat
ketosis and for use in formulated electrolyte treatments.

32 Glucose is one of several materials produced through the biological or chemical breakdown of starch. Each of 33 these materials is distinguished by the degree of starch hydrolysis, as well as by name and by CAS number. The 34 term "glucose" in this report refers to refined dextrorotatory¹ glucose (D-glucose), though it is known in the 35 glucose syrup industry as "dextrose" (BeMiller, 2009). Dextrose monohydrate is purified, crystalline D-glucose 36 containing one molecule of water of crystallization per molecule of D-glucose, and anhydrous dextrose is 37 purified, crystalline D-glucose without water of crystallization (BeMiller, 2009). Commercially, the term 'glucose" can also refer to glucose syrups (e.g., CAS# 8029-43-4) or corn syrups. These products are not the same 38 39 as refined D-glucose. Glucose syrups consist of a mixture of saccharides that result from incomplete hydrolysis of 40 starch (BeMiller, 2009; Jackson, 1995). Another product of starch hydrolysis is maltodextrin (CAS# 9050-36-6). 41 These related materials are not considered synonymous with glucose but may be discussed at times in this 42 technical report. 43

This report serves to provide technical information to complement the 1995 Technical Advisory Panel Report on glucose for the National Organic Standards Board (NOSB) to support the sunset review of glucose listed at 7 CFR §205.603(a)(13).

47 48

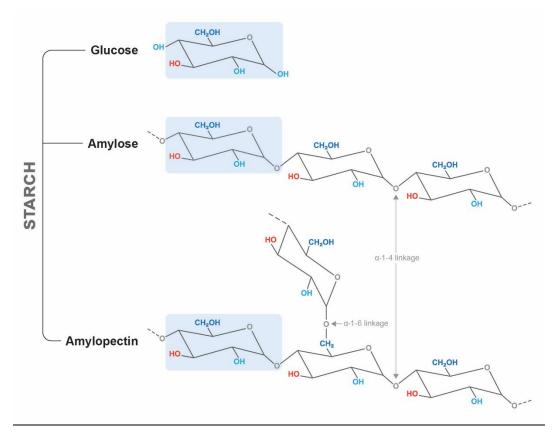
49

Characterization of Petitioned Substance

- 50 Composition of the Substance:
- 51 Glucose (also known as dextrose) is a 6-carbon (hexose) sugar molecule, and is the primary sugar in most
- 52 fruits and berries (Schenck, 2000). Glucose, like most other molecules, can exist in left and right-handed
- 53 versions, called enantiomers (Chemistry LibreTexts, 2015). D-glucose (subsequently referred to simply as
- ⁵⁴ "glucose") occurs naturally and is used by living organisms as the primary source of energy for cellular

¹ A compound is dextrorotatory when it is capable of rotating polarized light in the clockwise direction (Chang, 2000).

- respiration (Murphy et al., 2014). Glucose is often found naturally as a polymerized chain in materials such
- as cellulose, starch, and other carbohydrates (see Figure 1, below). Glucose can bond to other glucose
 molecules in different orientations. Some bond orientations are referred to as "alpha" (*α*), while others are
- ⁵⁷ "beta" (β). When glucose is connected via α -1,4 bonds,² it forms amylose (Murphy et al., 2014).
- 59 Amylopectin is formed when additional glucose molecules are attached via α -1,6 bonds (Murphy et al.,
- 60 2014). These molecules are the two main components of starch (Kearsley & Dziedzic, 1995; Murphy et al.,
- 61 2014; Schenck, 2000). If the hexose rings are connected through β -linkages, cellulose is the resulting polymer
- 62 (Schenck, 2000).
- 63



64

Figure 1: Chemical structure of glucose (A). Diagram also shows the two main components of starch: α -1-4 linked glucose polymers (amylose) (B) and α -1-4 and α -1-6 linked glucose polymers (amylopectin) (C). Illustration modified from Muralikrishna & Nirmala (2005).

65

66 Source or Origin of the Substance:

67 Glucose is commercially produced through the hydrolysis of starches, most commonly from maize

- 68 (Jackson, 1995; Olsen, 1995; Schenck, 2000). Other sources of starch may include wheat, rice, potato, barley,
- 69 sago and sorghum, depending on the global production location (Schenck, 2000; Zainab et al., 2011). The
- 70 hydrolysis catalysts are typically enzymes, but also include acids (BeMiller, 2009; Jackson, 1995; Olsen,
- 71 1995; Schenck, 2000).
- 72
- As mentioned, maize (*Zea mays* L.) is the major starch source worldwide, representing about 85% of
- vorldwide starch production (R. Zhang et al., 2021). United States is the biggest corn producer worldwide,
- vith a productive volume of over 345 million metric tons in 2019 (Shahbandeh, 2021). In the United States,
- the production of glucose from corn starch increased from 483,000 tons in 1964 to 642,000 tons in 1992 and
- reached 713,000 tons in 2019 (USDA-Economic Research Service, 2020).
- 78

 $^{^{2}}$ A bond is called " α -1,4 bond" when the α -hydroxyl functional group (-OH *below* the glucose ring) of the Carbon 1 (C1) of a glucose molecule bonds the α -hydroxyl functional group of the Carbon 4 (C4) of another glucose molecule, producing water and creating an O-glyosidic (oxygen mediated) link.

- 79 The degree of hydrolysis of the starch is commonly defined as the dextrose equivalent (DE). Complete
- hydrolysis of starch gives nearly pure glucose syrups or liquors. Crystalline glucose is produced from these
 highly refined glucose (94-95%+) liquors (BeMiller, 2009; Schenck, 2000). The liquor is refined by
- adsorption-separation chromatography,³ demineralization, evaporation and then finally crystallization to
- obtain either anhydrous dextrose (D-glucose) or dextrose monohydrate (D-glucose monohydrate)
- 84 (BeMiller, 2009), both of which are referred to as 'glucose' throughout this report.
- 85
- 86 See *Evaluation Question* #2 for details regarding specific glucose manufacturing processes.
- 87

88 **Properties of the Substance:**

- 89 Glucose is odorless and sweet, and soluble or miscible with water (National Center of Biotechnology, 2021).
- 90 Glucose injectable solutions are available at different concentrations (e.g. 5%-50% glucose anhydrous
- 91 and/or glucose monohydrate) (FDA, 2021). Table 1 summarizes some of the chemical and physical
- 92 properties of glucose.
- 93
- 94 95

Table 1: Properties of glucose

Property	Value
Physical State and Appearance	Crystalline powder (<i>a</i> -D-Glucose) ^b
Odor	Odorless ^a
Taste	Sweet ^a
Color	White ^a
Molecular Formula	$C_6H_{12}O_6^{b}$
Molecular Weight	180.16 ^b
Specific Gravity	1.56 ^b
pН	A 0.5 molar aqueous solution = 5.9 b
Solubility	Soluble ^b
рКа	12.92 at 0 °C ^b
Boiling Point	Greater than 212 °F at 760 mm Hg ^b
Melting Point	Less than 32 °F ^b
Critical Temperature	755 deg K (est) ^b
Vapor Pressure	8.0 x 10 ⁻¹⁴ mm Hg at 25 °C /extrapolated from a higher solid-phase temperature range ^b
Stability	Stable under proper storage conditions ^b
Reactivity	Weak reducing agent ^b

97 98

99 <u>Specific Uses of the Substance:</u>

100 Glucose, in solution and in its crystalline form, is primarily used for food and pharmaceutical purposes in

Source: a=(Schenck, 2000; Wilson et al., 1995), b=(National Center of Biotechnology, 2021)

101 the United States (Hull, 2010; Jackson, 1995; Macrae et al., 1993; Schenck, 2006). Glucose is used to treat

⁹⁶ 97

³ Adsorption-separation chromatography is a technology used to separate two substances using the different affinity they have for a resin. When a sample passes through columns that contain the resin, the rate of diffusion of the components causes them to separate as they flow through it (Coskun, 2016; Purolite, 2021).

Glucose

- metabolic disorders such as hypoglycemia⁴ (National Center of Biotechnology, 2021), as a component of
 certain products (e.g., electrolytes) and as an excipient (e.g., as a binder in oral tablets).
- 104105 Glucose is included at §205.603(a)(13) without annotation where its use is only restricted to medical
- 106 treatments (as well as preventive management standards per 7 CFR §205.238). While it is allowed for
- 107 livestock medical treatments beyond ketosis and dehydration, these two uses of the substance are the most
- 108 common. Glucose is also a common component of electrolyte formulations, and is used as an excipient in 109 livestock health care treatments
- 110
- 111 *Ketosis treatment*
- 112 One of the primary uses of glucose in organic production is in the treatment of ketosis in ruminants.
- 113 Ketosis is a metabolic disease that can occur shortly after parturition (labor and delivery) in ruminants due
- to an energy imbalance⁵ related to the sudden onset of milk production (Duffield, 2000; Herdt, 2000). It is
- fatal if untreated. Subclinical ketosis is defined as an increase of ketone bodies⁶ in the blood, urine, or milk,
 in absence of obvious clinical signs of disease (G. Zhang & Ametaj, 2020). Its primary feature is elevated
- 117 levels of ketones in the animal's blood stream (Andersson, 1988; Duffield, 2000). Clinical ketosis also
- presents elevated levels of ketones; in addition it includes loss of appetite, decreased milk production, and
- 119 loss of body condition (David Baird, 1982; Herdt, 2000). Both clinical and subclinical ketosis are also
- 120 associated with increased levels of non-esterified fatty acids (NEFA) and decreased levels of blood glucose
- 121 (Herdt, 2000; Mann et al., 2017). The hypoglycemic hypothesis states that alterations of glucose and lipid
- metabolism are associated with the development of ketosis, decreased level of glucose being one of the main diameter (2, 71) and (2, 71)
- major changes in affected animals (G. Zhang & Ametaj, 2020). Hypoglycemia can occur when the liver is not able to produce enough glucose to meet the demands of the postpartum ruminant. Low concentrations
- 125 of blood glucose are associated with low concentrations of insulin (hypoinsulinemia), which triggers the
- mobilization of fatty acids from adipose tissue (lipolysis), thereby increasing ketone body formation (G.
- 127 Zhang & Ametaj, 2020). Excessive lipolysis can lead to ketosis (G. Zhang & Ametaj, 2020). Ketosis can be
- monitored by measuring the amount of β -hydroxybutyrate (BHB), a ketone containing molecule, in the
- animal's blood (Gerloff, 2000; Gordon et al., 2013). Glucose is often given to ruminants through an
- 130 intravenous injection to replace the depleted, naturally occurring blood glucose. The replacement glucose
- serves as an energy supplement when the animal experiences negative energy balance and the nutritional
- demand of producing milk outstrips the dry matter intake the animal consumes (Herdt, 2000; Mann et al.,2017).
- 134
- 135 Ketosis is also discussed in detail within the 2021 *Propylene Glycol* Technical Report (USDA, 2021).
- 136
- 137 Neonatal hypoglycemia treatment
- 138 Immature neonates and neonate ruminants can become hypoglycemic because of underdeveloped
- 139 gluconeogenic mechanisms, if they do not ingest adequate amounts of colostrum and milk (Klein et al.,
- 140 2002). In cases of neonatal hypoglycemia, the immediate treatment consists of the intravenous or
- 141 intraperitoneal administration of a glucose solution. Under-nurtured neonatal calves and
- 142 immunosuppressed animals are predisposed to coliseptisemia (invasion of the blood stream by coliform
- bacteria). Animals affected by this disease are usually treated by the intravenous administration of large
- volumes of balanced electrolyte solutions over several hours; fluids should include glucose to correct
- 145 hypoglycemia (Walter, 2020).
- 146
- 147 Formulated oral electrolyte solutions and rehydration therapies
- 148 Glucose helps facilitate sodium transport within the intestines (Naylor, 1990). Because of this, it is a key
- 149 ingredient in oral rehydration therapies to treat dehydration in young ruminants. Calves, lambs, kids, and
- 150 swine are most likely to benefit from oral electrolyte solutions. Neonatal diarrhea (scours) remains the most
- 151 common cause of death in beef and dairy calves (Smith, 2009). Young livestock often experience

 $^{^{\}rm 4}$ An abnormally diminished content of glucose in the blood (Rozance & Hay, 2010).

⁵ An imbalance between the energy that enters into the body as feed (dry-matter intake) and the energy that is released from the body in the form of milk (G. Zhang & Ametaj, 2020).

⁶ ketone bodies are hydroxybutyrate (OHB), acetoacetate (AcAc), and acetone (Ac) and can be found in the blood, urine, and milk of cows in ketosis (G. Zhang & Ametaj, 2020).

dehydration due to diarrhea following an infection by *E. coli* or cryptosporidium (Naylor, 1999). This

153 causes the animals to expel (rather than absorb) the large amounts of fluid that is secreted in the small 154 intestine. Regardless of the pathogen and mechanism involved, diarrhea increases the loss of electrolytes

and water in the feces of calves, and decreases milk intake, resulting in dehydration and negative energy

156 balance (Smith, 2009). Diarrhea is by far the most common indication for fluid therapy in neonatal calves.

157 Oral electrolyte solutions have classically been used to replace fluid losses, correct acid-base and electrolyte

abnormalities, and provide nutritional support (Smith, 2009).

159

160 Oral electrolyte solutions were developed in the twentieth century as a treatment for cholera infections.

161 The original World Health Organization (WHO) electrolyte was based on a formulation that contained an

approximately equimolar mixture of sodium (990 mmol/ L) and glucose (2%), potassium, glycine and

bicarbonate (Smith, 2009). Although much research has been done on oral fluid therapy since that time, the

164 formulation of oral fluids has not moved far from the original (Smith, 2009). Commonly recommended oral 165 robudration solutions contain 75 mmol to 120 mmol (L of glucose (Boid & Local, 2000)

rehydration solutions contain 75 mmol to 139 mmol/L of glucose (Reid & Losek, 2009).

167 Excipient

168 Glucose is a common excipient ingredient in livestock health care products (OMRI, 2021), and meets the

annotation for excipients used in drugs and biologics used to treat organic livestock at §205.603(f)(1).

170171 *Other uses*

As an ingredient, glucose is generally recognized as safe (GRAS) by the U.S. FDA (21 CFR 184.1857)

173 without limitation when used in food. Glucose monohydrate (usually referred to as dextrose monohydrate)

174 is highly valued as an ingredient in confectionery applications (Jackson, 1995). It is important for preserves.

175 At any given concentration, a dextrose solution contains almost twice as many dissolved molecules as a

176 sucrose solution, and therefore a solution of glucose exerts a greater osmotic pressure than a sucrose

solution (Jackson, 1995), aiding with the osmotic dehydration. Glucose is also a valuable ingredient in

178 powdered sherbet centers, lemonade powders, chewing gum, compressed tablets and fondant (Jackson,

179 1995). It is also sometimes used in brewing (Schenck, 2000).

180

181 Glucose is used in the production of microbially-derived products, such as citric, lactic, and acetic acids, as

182 well as enzymes, vitamin C, and antibiotics (Schenck, 2000). It is also used in producing fuel ethanol,

183 plastics, insulating foam, and adhesives (Schenck, 2000). Glucose is used in conventional livestock feeds as

an appetite stimulant due to its sweet flavor (Precision Feed Technologies, LLC, 2021; Stock Show Secrets,

- 185 2022; Aspen Veterinary Resources, Ltd., 2021)..
- 186

187 Anhydrous glucose (anhydrous dextrose) is used for intravenous injections in humans for various

pharmaceutical and medicinal preparation (Fellers, 1939). In the pharmaceutical industry, glucose is found

as both an active ingredient, and as an excipient (inactive ingredient). Glucose injectable solutions are used

as a source of water and calories for patients that required intravenous nutrition (FDA, 2021). As an

191 excipient, glucose has widespread use as a sweetener, reducing agent, bulking agent and soluble carrier for

- an active pharmaceutical ingredient (Srivastava et al., 2016).
- 192 an active pharmaceutical ingredient (Srivastava et al., 2016).193
- 193

195 Approved Legal Uses of the Substance:

196 Food and Drug Administration (FDA)

D-glucose appears in the "Corn sugar" listing at 21 CFR 184.1857. It is considered a substance that, when
added directly to human food, is generally recognized as safe (GRAS). Glucose sirup (also spelled "syrup")

199 is found under the "Corn Syrup" listing (21 CFR 184.1865) and under the sweeteners and table sirups

- section (21 CFR 168.120). Corn sugar and corn syrup are allowed as food ingredients with no limitation
- 201 other than current good manufacturing practice; glucose syrup is allowed also as a sweetener and table
- 202 syrup (21 CFR 168.120).203

204 Environmental Protection Agency (EPA)

- 205 Dextrose and corn syrup appear on the 2004 EPA List 4A as inert ingredients of minimal risk (USA EPA,
- 206 2004). "D-glucose," "Corn syrup" and "syrups, corn, dehydrated" are also considered to fall under the

- category of "commodity inert", and are therefore approved for food and non-food pesticidal use as inerts 207 208 (US EPA, 2004).
- 209

210 Action of the Substance:

211 Hypoglycemia and ketosis treatment

When delivered intravenously, glucose provides an immediate supply of sugars to the blood stream and 212

213 effectively treats nervous ketosis,⁷ the most severe form of the disease (Gordon et al., 2013). Because

214 glucose is immediately bioavailable to ruminants, its effects are not long-lasting (Wagner & Schimek, 2010).

215 Glucose provides less than 12 hours of suppression of BHB, a ketone often used as a marker for ketosis,

- 216 and only one treatment of 500 mL or 1 L of 50 percent glucose is unlikely to prevent or resolve ketosis in a
- 217 dairy cow (Wagner & Schimek, 2010). Dairy cows may need follow-up treatment when using glucose
- 218 because each dose is effective for less than 12 hours (Herdt & Emery, 1992). Oral administration of glucose
- 219 to sheep is possible, but research suggests that sheep may not successfully absorb the needed amount of 220 glucose through their rumen (Sargison, 2007).
- 221

222 Gordon et al. 2013 observes that dextrose (glucose) should be considered a second-line treatment for cases

- 223 of ketosis. The treatment with dextrose should be used in animals with severe ketonemia and concurrent
- 224 hypoglycemia suffering from nervous signs (abnormal licking, chewing on pipes or concrete, gait
- 225 abnormalities, and aggression). These animals should then receive additional other treatments for longer-
- 226 term effectiveness (Gordon et al., 2013).
- 227
- 228 Dehydration treatment
- 229 Glucose can be co-transported with sodium from the intestinal lumen to the inside of the enterocyte at the
- 230 brush border membrane (special epithelium found in some tissues, like the intestine) (Smith, 2009). At the
- 231 basolateral membrane, specific transmembrane enzymes actively pump sodium ions out of the cell, thus
- raising the intercellular osmolality (Smith, 2009). This increase in intercellular osmolality then draws more 232
- 233 water from the intestinal lumen through the tight junctions between cells, thereby expanding extracellular
- 234 fluid volume and rehydrating the animal (Smith, 2009).
- 235

236 **Combinations of the Substance:**

- 237 Glucose is commercially available in two forms – in diluted liquid solutions and in a crystalline powder.
- 238 Some products contain hydrochloric acid or sodium hydroxide for pH adjustment (VetOne®, 2022). Aside 239 from that, intravenous dextrose (glucose) is formulated with sterile water.
- 240

241 In the case of oral electrolytes, some products may be formulated with glucose, certain salts in the form of 242 ions - sodium, potassium, chloride, acetate, citrate, etc., and amino acids to aid with the hydration process. 243 Sometimes preservatives like citric acid or propionic acid are included as part of the formulation. Other

- 244 substances may be added to enrich products and improve the nutritional intake of the treated livestock. For
- 245 example, some products may be enriched with vitamins, microorganisms, and/or amino acids like glycine
- 246 (Agri Laboratories, Ltd., 2022). Whether amino acids are needed in addition to glucose in oral electrolyte
- solutions is not well understood; however, the addition of glycine does seem to further improve water 247
- 248 absorption in the intestine (Smith, 2009).
- 249 250

251

Status

252 **Historic Use:**

- 253 Ketosis treatment
- 254 Ketosis as a disorder in cattle has been known since at least 1849 (McSherryt et al., 1960). In 1928, Hupka
- 255 noted that administering glucose helped alleviate symptoms of ketosis (McSherryt et al., 1960). Since the
- 256 1930s, glucose has been considered a staple to treat hypoglycemia associated with ketosis (Gordon et al., 2013).
- 257

²⁵⁸

⁷ Nervous ketosis is marked by signs that may include excitement and hyperesthesia, depraved chewing and licking (occasionally with self-mutilation), or abnormal gait (including hypermetria or ataxia) (Gerloff, 2000).

- Glucose was studied extensively in the 1940s and 1950s, but researchers relied on studies where all affected
- animals were given the same treatment, and no controls were used for comparison (Gordon et al., 2013).
- According to Gordon et al., as of 2013, glucose has never been studied in a randomized clinical trial to
- 262 determine efficacy as a standard treatment for ketosis.
- 263
- Mann et al. (2013) found that treating cattle with a combination of glucose and propylene glycol reduced
- BHB more than either substance alone. Capel et al. (2021) also investigated the effect of intravenous glucose
- treatment combined with oral propylene glycol therapy on the resolution of lactating cow's
- 267 hyperketonemia by assessing the levels of blood BHB. In contrast to what Mann et al. found, the addition
- of glucose for 1 to 3 days provided no improvement in resolution of ketosis.
- 269
- 270 *Dehydration treatment*
- 271 Oral electrolyte solutions became widely commercially available in the early 1970s and gained rapid
- acceptance in the treatment of diarrheic animals (Naylor, 1990). Oral electrolytes continue to be the
- hallmark of routine therapy for treating neonatal calf diarrhea (Smith, 2009). Glucose is present in various
- concentrations in virtually all commercially available oral electrolyte solutions (Smith, 2009). In addition to
- the treatment of sick neonatal calves, fluids and electrolytes are used in sick ruminants to correct
- 276 imbalances of acid-base, electrolyte, or water, and to optimize tissue blood flow, provide nutrients, or treat
- 277 shock (Constable, 2003).
- 278
 279 Organic Foods Production Act, USDA Final Rule:
- 280 The NOSB recommended including glucose on the National List in 1995 as a synthetic material allowed for
- 281 use in livestock medical treatments. The National Organic Program currently allows glucose at 7 CFR
- 282 §205.603(a)(13) for use as a disinfectant, sanitizer, and medical treatment as applicable. As a medical
- treatment, glucose is limited to use after the onset of illness by 7 CFR 205.238(c)(2).
- 284
- 285 <u>International</u>
- 286 Canadian General Standards Board Permitted Substances List
- 287 The Canadian General Standards Board includes glucose on CAN/CGSB 32.311-2020 Table 5.3 (Health
- 288 Care Products and Production Aids) without annotation. Table 5.3 also includes a listing for "Formulants"
- 289 (inerts, excipients)," allowing glucose to be used as an excipient ingredient with a permitted active
- 290 ingredient. CAN/CGSB 32.310-2020 6.6.2 prohibits the use of veterinary drugs in the absence of illness.
- 291
- 292 CODEX Alimentarius Commission, Guidelines for the Production, Processing, Labelling and Marketing of
 293 Organically Produced Foods (GL 32-1999)
- 294 The CODEX guidelines state in Annex 1, Part B "Health Care" that producers must first prevent disease
- through the selection of appropriate breeds, use of high-quality feed, and access to pasture and exercise,
- among other preventive principles. If these management practices are not enough to prevent disease, a
- 297 producer may use allopathic⁸ veterinary drugs if phytotherapeutic products are ineffective. Glucose is not
- explicitly mentioned as a health care substance, but it is included in allopathic veterinary drugs and
- therefore allowed based on the definition at Section 2.2 of the guidelines.
- 300
- 301 European Economic Community (EEC) Council Regulation, EC No. 834/2007 and 889/2008
- 302 Title II, Chapter 2, Section 4 of the EC No. 889/2008 focuses on disease prevention and veterinary
- 303 treatment. Article 24, paragraph 3 requires producers to use preventive measure to ensure animal health,
- and also allows producers to use veterinary medicinal products if prevention or phytotherapeutic products
- fail. Glucose is included in veterinary medicinal products, and therefore would be allowed under EEC
- 306 regulations. Article 24, paragraph 5 requires that organically produced foodstuffs from treated animals be
- 307 withheld from the stream of commerce for twice the legal withdrawal period or at least 48 hours.
- 308
- 309 Japan Agricultural Standard (JAS) for Organic Production

⁸ Allopathy was coined in 1810 by German physician Samuel Hahmann to designate the usual (western) practice of medicine. Allopathic medicine focuses on signs and symptoms of the diseases, identifying the pathology behind the disease and treating them with drugs, surgery, etc. (Parajuli & Sanjib, 2021).

Technical Evaluation Report

Glucose

310 Article 4 of the Japanese Agricultural Standard for Organic Livestock includes the "Health control" section, 311 specifying practices for organic livestock production. The standard requires that producers implement 312 preventive practices before using veterinary drugs, and veterinary drugs may only be used for therapeutic 313 purposes. Again, glucose is included within veterinary drugs and may therefore be allowed under the JAS. 314 A withdrawal period is noted. It must be 48 hours from the last administration of drugs to slaughter for 315 foods, milking, and egg collection, or twice the period of drug withdrawal defined by Articles 14-1, 9, 4, 316 and 6 of the Pharmaceutical Law for the approval of drugs, change of approvals, reexamination of drugs, 317 and drug efficacy review, whichever is longer. 318 319 IFOAM – Organics International 320 Section 5.6 of the IFOAM Standard for Organic Production and Processing describes the requirements for 321 the use of veterinary medicine in organic livestock production. Section 5.6.1 requires that producers 322 establish preventive practices, including good quality feed and access to the outdoors, to avoid illness in their livestock before using synthetic allopathic veterinary medical products. Glucose, when used to 323 324 address dehydration and ketosis symptoms in livestock, would be considered a synthetic allopathic 325 veterinary medical product, and Exception (c) would allow its use under veterinary supervision with a 326 minimum withdrawal period of at least 14 days. Prophylactic use of synthetic allopathic veterinary drugs 327 is prohibited. 328 329 330 331 Evaluation Questions for Substances to be used in Organic Crop or Livestock Production 332 333 Evaluation Question #1: Indicate which category in OFPA that the substance falls under: (A) Does the 334 substance contain an active ingredient in any of the following categories: copper and sulfur 335 compounds, toxins derived from bacteria; pheromones, soaps, horticultural oils, fish emulsions, treated 336 seed, vitamins and minerals; livestock parasiticides and medicines and production aids including 337 netting, tree wraps and seals, insect traps, sticky barriers, row covers, and equipment cleansers? (B) Is the substance a synthetic inert ingredient that is not classified by the EPA as inerts of toxicological 338 339 concern (i.e., EPA List 4 inerts) (7 U.S.C. § 6517(c)(1)(B)(ii))? Is the synthetic substance an inert 340 ingredient which is not on EPA List 4, but is exempt from a requirement of a tolerance, per 40 CFR part 341 180? 342 343 (A) Glucose is used as an active ingredient in livestock medicines. 344 (B) Glucose is a 2004 EPA List 4A inert ingredient of minimal risk (US EPA, 2004). Furthermore, dextrose 345 (D-glucose; CAS No. 50-99-7) and dextrose monohydrate (D-glucose monohydrate; CAS No. 77938-63-7) are also considered "commodity inerts" that are exempt from the requirement of a tolerance at 40 CFR 346 347 §180.950(a)(i) (US EPA, 2020). 348 349 Evaluation Question #2: Describe the most prevalent processes used to manufacture or formulate the 350 petitioned substance. Further, describe any chemical change that may occur during manufacture or 351 formulation of the petitioned substance when this substance is extracted from naturally occurring plant, 352 animal, or mineral sources (7 U.S.C. § 6502 (21)). 353 354 Manufacturers produce glucose using these basic chemical and physical steps: hydrolysis, clarification, color removal, evaporation, and crystallization (for crystalline dextrose, or glucose, production). Currently, 355 manufacturers produce crystalline glucose products from acid-enzyme and enzyme-hydrolyzed glucose 356 syrups (Schenck, 2000). In the past, glucose was obtained from acid-hydrolyzed syrups. 357 358 Corn starch slurry obtained through the wet milling process is usually the starting material for glucose production. The pH of the starch slurry is adjusted to 6.0 and calcium ions (usually in the form of calcium 359 360 chloride) may be added in order to stabilize or improve the efficiency of the enzymes (Schenck, 2000).

[Insert date report is transmitted to NOP]

- When preparing the initial starch slurry, sulfur dioxide may be added in order to minimize bacterial colonization and to block the Maillard reaction (the reaction of proteins with reducing sugars, which
- 363 produces colors) (Hull, 2010).
- 364 *Enzymatic hydrolysis*:
- 365 Using the enzymatic hydrolysis process, starch is hydrolyzed by the addition of high-temperature stable
- 366 enzymes. Using direct steam injection or some other method, the starch slurry with enzymes is heated to
- 367 approximately 105 °C in order to liquefy it (Schenck, 2000). The pH and other conditions might be
- readjusted, and more enzyme(s) may be added for starch hydrolysis (saccharification) (Hull, 2010; Macrae
- 369 et al., 1993; Olsen, 1995).
- To process starch into glucose, four main type of enzymes are used: α-amylase, β-amylase, glucoamylase
- and pullulanase (BeMiller, 2009). The enzyme α-amylase is one of the most important enzymes used in the
- food industry, and these enzymes account for approximately 25% of the world enzyme market (de Souza
- 373& Oliveira Magalhães, 2010). They hydrolyze the α-1,4-glycosidic linkages of the starch polysaccharide.
- Thermostable (heat-stable) α-amylases are desirable because liquefaction and saccharification of starch are
- 375 performed at high temperatures (de Souza & Oliveira Magalhães, 2010; Hull, 2010; Reddy et al., 2003).
- 376 Industrially, saccharification is predominantly carried out by α-amylases from *Bacillus amyloliquefaciens*,
- 377 *Bacillus licheniformis*, or *Bacillus stearothermophilus* (Bilal & Iqbal, 2020). The commercial α-amylases are
- 378 produced by fermentations of genetically modified bacteria, where the native gene has been manipulated
- to code for an enzyme with improved performance characteristics, such as heat stability (Nielsen, 2012;
- 380 University of Reading NCBE, 2018; DuPont Industrial Biosciences, 2015; Olempska-Beer, 2004; Silano et al.,
- 381 2018).
- 382 *Acid hydrolysis:*
- Acid hydrolysis may be used to partially hydrolyze the starch slurry before further enzyme hydrolysis, as
- 384 well as to make 35 and 42 DE (dextrose equivalent) finished glucose syrups. In the acid conversion process,
- a starch slurry is acidified (usually with hydrochloric acid) to a pH of about 2 and pumped to a vessel
- 386 where it is heated and pressurized. This process partially hydrolyzes the starch slurry. After hydrolysis, the
- slurry is neutralized (usually with sodium carbonate) to a pH 4.5-4.8, which causes proteins and lipids to
- 388 precipitate.
- 389 When the end product is a glucose syrup, the slurry is then purified by centrifugation, skimming and/or
- passing through deep tanks to remove impurities (solids, fats, proteins, oils and fine fibers), and then
- filtered. The product is clarified using granular activated carbon to further remove impurities, and then
- 392 concentrated by evaporation. The resulting syrup is polished through further clarification and
- decolorization. Finally, the syrup is concentrated again in evaporators to the final required density. Some
- 394 syrups are treated with ion exchange resins for further refinement (Hull, 2010; Macrae et al., 1993;
- 395 Mironescu & Mironescu, 2006).
- 396 Acid–enzyme process:
- 397 The combination of acid and enzymes is used to produce high glucose syrups such as D.95 (95% glucose).
- 398 The starch slurry is only partially converted by acid to a given DE The temperature and acidity of the
- slurry are adjusted to the optimal conditions required by the specific enzyme or enzymes to be used during
- the saccharification process, where the starch is broken down into monosaccharides. The DE is monitored
- and the conversion processes stopped when the desired sugar composition is reached. The syrup is
- 402 centrifuged, filtered, clarified with activated carbon and ion exchange treatment, polished, and evaporated
- 403 as needed (Hull, 2010; Macrae et al., 1993; Olsen, 1995). Glucoamylase (also known as amyloglucosidase or
- 404 AMG) is used after the acid hydrolysis and the conversion is mediated by α-amylase (Hull, 2010).
- 405
- 406 *Purification:*
- Glucose monohydrate crystals are produced through the crystallization of 95 DE syrups inside crystallizers (large horizontal, cylindrical batch tanks) (BeMiller, 2009). Inside these tanks, the syrup is cooled to achieve

- 409 the proper level of supersaturation (temperature and concentration conditions required for the crystals to 410 precipitate), and subsequently to achieve the crystallization of glucose in its monohydrated form (BeMiller, 2009). The crop of crystals is then washed and centrifuged in basket centrifuges to remove the remaining 411 412 liquor, which may be reprocessed to yield a second crop of crystals (BeMiller, 2009). Crystals are then dried 413 with a stream of hot air, cooled, and stored. Throughout this purification process a product containing 414 99.9% dextrose can be obtained (Hull, 2010). Anhydrous dextrose is produced by dissolving the 415 monohydrate in hot purified water and refining it again (BeMiller, 2009). 416 417 During the purification and crystallization steps to make glucose, the enzymes are typically removed. 418 Absence of the α-amylase protein in the final (purified) sweetener syrup has been confirmed 419 experimentally (Pronk & Leclercq, 2004). In addition, governmental and international organizations such as the FDA and the World Health Organization (WHO) through the International Program on Chemical 420 421 Safety (IPCS) have observed that these enzymes are GRAS and Allowable Daily Intake (ADI) not specified, 422 respectively (Pronk & Leclercq, 2004). 423 424 Evaluation Question #3: Discuss whether the petitioned substance is formulated or manufactured by a 425 chemical process, or created by naturally occurring biological processes (7 U.S.C. § 6502 (21)). 426 427 Glucose is produced naturally through photosynthesis and is stored in plants in the form of starch, a 428 polymer made from glucose. Current industrial processes use a combination of biological, chemical and physical tools to obtain purified glucose. These processes yield products that can be $\geq 99.5\%$ pure. 429 430 Glucose is classified⁹ as a synthetic substance when acid hydrolysis and acid/enzyme hydrolysis are used 431 in the production, but as a nonsynthetic material when the production is achieved through enzyme hydrolysis.
- 432

433

- 434 Commercially available products using glucose can be formulated with synthetic substances. Some
- 435 products such as intravenous dextrose solutions are diluted in water to achieved desired concentrations.
- 436 These injectable solutions may contain hydrochloric acid or sodium hydroxide for pH adjustment. In the
- 437 case of oral electrolytes, these types of products may be formulated with glucose, certain salts in the form
- of ions sodium, magnesium, etc., and amino acids to aid with the hydration process, and sometimes 438
- 439 preservatives like citric acid or propionic acid are included as part of the formulation.
- 440

441 Evaluation Question #4: Describe the persistence or concentration of the petitioned substance and/or its 442 by-products in the environment (7 U.S.C. § 6518 (m) (2)).

443

444 Glucose and glucose-containing compounds are naturally abundant in the environment. Generally

- 445 speaking, sugars such as glucose are the most abundant organic compounds in the biosphere because they
- 446 are the basic components of all polysaccharides (chitin, cellulose, hemicellulose, starch, pectin, etc.)
- (Gunina & Kuzyakov, 2015). 447

448 Glucose is an easily metabolized substance (Brosnan, 1999; Murphy et al., 2014). Its use as an animal drug 449 is not expected to contribute to significant quantities in the environment. However, when glucose is given

- 450 to animals, some may be excreted in urine. For example, when glucose was given intravenously to healthy
- 451 cows, 13 to 26% of the glucose was excreted in the urine, depending on the total quantity given (Metzner et
- 452 al., 1993). Soil systems with active microbes should easily consume these amounts of glucose (in a matter of
- 453 hours to days, depending in the type of soil) if present in the excreted urine of treated animals as shown by
- 454 the studies performed by Ferreira et al. 2013, Padmanabhan et al. 2003 and Gunina et al. 2015. For more
- information regarding these studies, refer to Evaluation Question #8. 455
- 456

⁹ Considering the Decision Tree for Classification of Materials as Synthetic or Nonsynthetic in NOP Guidance 5033-1,

Evaluation Question #5: Describe the toxicity and mode of action of the substance and of its

458 breakdown products and any contaminants. Describe the persistence and areas of concentration in the 459 environment of the substance and its breakdown products (7 U.S.C. § 6518 (m) (2)). 460 Glucose is an important biomolecule that has very low toxicity. According to several safety data sheets, the oral LD₅₀ in rats is 25800 mg/kg (DHI Milieu Ltd., 2008; Fisher Science Education, 2015; Hach Company, 461 2005). When glucose is metabolized through aerobic respiration, the breakdown products are water and 462 CO_2 (Murphy et al., 2014). Plants and cyanobacteria recycle CO_2 and water back into glucose via 463 photosynthesis (Galant et al., 2015). Glucose typically persists in the environment within polymers such as 464 chitin and cellulose (Gunina & Kuzyakov, 2015). 465 466 Evaluation Ouestion #6: Describe any environmental contamination that could result from the 467 468 petitioned substance's manufacture, use, misuse, or disposal (7 U.S.C. § 6518 (m) (3)). 469 470 The major environmental impacts associated with the production of starch, from which glucose is derived, 471 occur during the agricultural stages that produce the starch-containing material used for glucose 472 production (e.g., potato, maize, wheat, and cassava) (Blanco-Cejas et al., 2020). The agricultural stages 473 usually involve intensive consumption of natural resources such as land occupation and transformation, 474 use of fertilizers and pesticides, depletion of fossil fuels for machinery, etc. (Blanco-Cejas et al., 2020). 475 476 The results of a life cycle assessment¹⁰ (LCA) for glucose ascribe 60-96% of the generated impact to the 477 production of the starch (Blanco-Cejas et al., 2020). Previous studies noted that 70% of the environmental 478 footprint of glucose production is generated by the starch manufacture (Blanco-Cejas et al., 2020). 479 480 The impact to produce 1000 kg of glucose from corn (100% DM) was quantified by Renoulf et al. (2008) at 481 6000 MJ of energy input; 1000 kg CO_{2eq} for global warming; 8.5 kg SO_{4eq} for acidification and 2.8 kg PO_{4eq} 482 for eutrophication potential (Kis et al., 2019; Renouf et al., 2008). By comparison, Renoulf et al. (2008) note 483 that glucose from sugar cane is more sustainable than corn-derived glucose in terms of energy input, greenhouse gas emissions and possibly acidification potential. Kis et al. 2019 noted that inverted liquid 484 485 sugar has lower carbon and water footprints than glucose and fructose syrups and derivatives (by 38% and 486 95%, respectively), and its production requires less fossil energy (by 31%) and less agricultural land (by 487 67%). After evaluating an LCA for EU starch manufacturing plants, Vercalsteren et al. (2012) noted that the 488 starch industry typically causes little waste production because all side streams are used to produce useful 489 products that have an economic value; waste sent to landfill or incineration is almost nonexistent. Glucose 490 is one of the side stream products of the starch industry and, as a livestock medical treatment, it is unlikely 491 to contaminate the environment. Contamination related to the disposal of this product represents a 492 negligible risk to the environment. 493 494 Evaluation Question #7: Describe any known chemical interactions between the petitioned substance 495 and other substances used in organic crop or livestock production or handling. Describe any

496 environmental or human health effects from these chemical interactions (7 U.S.C. § 6518 (m) (1)).

497

457

498 Glucose may interact aggressively with strong oxidizing agents and can produce toxic gases when

499 combusted (Hach Company, 2005), however these reactions are unlikely to happen in the utility context of

500 livestock medicine. For environmental effects that the glucose may have, please review *Evaluation Question*

- 501 #4. For human health and glucose metabolism, please review *Evaluation Question* #3. Glucose is unlikely to
- 502 cause serious damage to producers.
- 503

504 <u>Evaluation Question #8:</u> Describe any effects of the petitioned substance on biological or chemical 505 interactions in the agro-ecosystem, including physiological effects on soil organisms (including the salt 506 index and solubility of the soil), crops, and livestock (7 U.S.C. § 6518 (m) (5)).

¹⁰ A life cycle assessment (LCA) is a method to analyze the environmental impacts of a product. An LCA quantifies the potential environmental effects of a product over its entire life cycle, meaning that the extraction of raw materials, the production of the materials and the product, the use and the end-of-life treatment are taken into account (Vercalsteren et al., 2012).

507

Glucose is a universal fuel for cellular metabolism. Glucose in the environment is captured rapidly by
microbes where it is used for maintenance and growth (Gunina & Kuzyakov, 2015). Glucose addition to the
soil has been utilized as a strategy for measuring the respiratory response of the soil microbial community
(Ferreira et al., 2013). Glucose labeled ¹⁴C and ¹³C are often used to perform biodegradation assays that
measure the microbial activity of water and soil samples. Soil microbes can mineralize¹¹ glucose added to

513 the soil (at 7 percent) within the first 8 hours of exposure (Padmanabhan et al., 2003).

514 Gunina & Kuzyakov (2015) estimated the sugar C mineralization to CO₂ using a literature review of 74 data

points collected from 16 studies on glucose ¹⁴C or ¹³C decomposition within the first 24 h after its addition

516 into the soil. The calculations performed showed that the estimated maximum glucose C decomposition

rate to CO_2 was 1.1 percent min⁻¹ (Gunina & Kuzyakov, 2015). At this high rate, half of the glucose C should be mineralized to CO_2 within the first hour (Gunina & Kuzyakov, 2015). This study also shows that

519 the time of glucose duration as a whole molecule during microbial metabolization is much shorter that 30

- 520 min (Gunina & Kuzyakov, 2015).
- 521

Ferreria et al. (2013) demonstrated that soil systems with no tillage use more carbon than systems with tillage, and that the microbes in these kinds of systems can consume up to 2000 mg of glucose kg⁻¹ dry soil

- 524 after 24 h of incubation.
- 525

526 In aquatic environments, glucose alone can support 20-30 percent of bacterial production in some oceanic

527 regimes and, as observed, in one Danish lake (Kirchman, 2003). In the Gulf of Mexico, Antarctic seas, and

in two Swedish lakes, glucose accounted for <10% of bacterial growth (Kirchman, 2003). In the surface

waters of the Gulf of Mexico, glucose was found at a concentration of 2-15 nmol/L (Skoog et al., 1999).

Gocke et al. (2003) found that turnover rates of glucose were very fast in highly productive lagoons: less

- than 20 minutes. In less productive systems, the cycling of glucose had a turnover time of two hours(Gocke et al., 2003).
- 532 (Gocke et al., 2 533

534 Considering the studies above, it is possible to conclude that glucose given to cows intravenously, as a 535 component of an electrolyte treatment, or as an excipient in other medical products, does not represent a

536 threat to water and soil systems. The glucose that is not metabolized might be excreted in the urine of the

537 treated animals, and the concentrations would be small enough that soil systems with active microbes

- should easily consume these amounts of glucose in a matter of hours to days depending in the type of soil.
- 539

540 <u>Evaluation Question #9:</u> Discuss and summarize findings on whether the use of the petitioned 541 substance may be harmful to the environment (7 U.S.C. § 6517 (c) (1) (A) (i) and 7 U.S.C. § 6517 (c) (2) (A) 542 (i)).

542 (1 543

544 As a substance that is critical to the metabolism of living cells, glucose is naturally pervasive in the

environment (Brosnan, 1999; Gunina & Kuzyakov, 2015). The use of glucose as intended at 7 CFR

- 546 §205.603(a)(13) by organic livestock producers is therefore unlikely to cause harm to the environment.
- 547 Manufacturing glucose does have the potential to cause environmental damage. The major environmental
- 548 impact of glucose production is associated with the agricultural production of the starch-containing
- produce (corn, wheat, potato, etc.) (Blanco-Cejas et al., 2020; Kis et al., 2019; Vercalsteren et al., 2012). About

550 70-96% of the ecological impact of glucose manufacturing is caused during the starch production (Blanco-

551 Cejas et al., 2020). However, if designed optimally, starch production plants should cause little waste

552 production because all the side streams can be used to produce economically valued products

- 553 (Vercalsteren et al., 2012).
- 554
- As described in previous sections, glucose is a universal energy source for living organisms. It is not

acutely toxic for animals, and almost any organism easily metabolizes it. Microorganisms in water and soil

557 decompose glucose into CO₂ and water. Plants and cyanobacteria take CO₂ and water and produce glucose

¹¹ The term mineralization is often used in microbial respiration studies and it describes the degradation of a compound to its "mineral components" (i.e. carbon dioxide and water) and is synonymous with ultimate biodegradation or complete biodegradation (Knapp & Bromley-Challoner, 2003).

- and other carbohydrates like starch, cellulose, and chitin through photosynthesis, closing the
- 559 biogeochemical cycle.
- 560

561Evaluation Question #10:Describe and summarize any reported effects upon human health from use of562the petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (i), 7 U.S.C. § 6517 (c) (2) (A) (i)) and 7 U.S.C. § 6518563(m) (4)).

564

565 High levels of glucose in the blood for a persistent period (hyperglycemia) can have a toxic effect on cells, tissues and organ systems (Giri et al., 2018). Insulin, secreted from the pancreatic β cells, is a key element in 566 the homeostatic regulation of blood glucose levels (Fujii et al., 2019). A prolonged hyperglycemic condition 567 568 leads to severe diabetic condition by damaging the pancreatic β -cell and inducing insulin resistance (Giri et al., 2018). People suffering from diabetes or prediabetes have a reduced ability to tolerate glucose loads, 569 570 and therefore their health could be negatively affected if they were to receive intravenous glucose 571 treatment unpaired with an insulin treatment (Dagogo-Jack & Alberti, 2002). Glucose as a component of 572 livestock health care products does not represent a health risk for the producers because they would not be 573 ingesting the substance. In addition, the glucose that is not metabolized by the animal will not persist in the 574 dairy products or meat, as it is excreted in the urine (Metzner et al., 1993).

- 575
- 576

577 <u>Evaluation Question #11:</u> Describe all natural (non-synthetic) substances or products which may be 578 used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed 579 substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).

580581 *Glucose*

582 Glucose production utilizing solely the enzyme hydrolysis process (Refer to *Evaluation Question* #2 for

further information) would yield a nonsynthetic product per NOP 5033-1 "Guidance: Decision Tree for
 Classification for Materials as Synthetic or Nonsynthetic".

- 585
- 586 Molasses

587 Molasses is a nonsynthetic, agricultural commodity commonly added to livestock feed. Adding molasses

as a top-dressing to forage (or fed directly as a fluid) can be used pre-partum as a preventive measure, and

- as a treatment for subclinical ketosis postpartum (Havekes et al., 2020; Lans et al., 2007). For more
- information, review *Evaluation Question* #11 of the 2021 Propylene Glycol Technical Report (2021).
- 591
- 592 Glycerin

Nonsynthetic glycerin, or glycerol, can be used for the treatment of ketosis. It can be delivered either as an

- oral drench or combined in the feed ration. Glycerin can be of special benefit to treat ketosis in sheep (Cal-
- 595 Pereyra et al., 2015; Ferraro et al., 2016; Kalyesubula et al., 2019). High dosages of glycerin may have
- negative impacts on biodiversity in the rumen, and work remains to clarify rumen impact of glycerin use
- 597 (Kupczyński et al., 2020).
- 598

599 <u>Evaluation Question #12:</u> Describe any alternative practices that would make the use of the petitioned 600 substance unnecessary (7 U.S.C. § 6518 (m) (6)).

- 601
- 602 Ketosis:

603 Several studies have found that animals that are given the opportunity to graze and eat high-forage diets 604 have a decreased incidence of ketosis (Richert et al., 2013; Vickers et al., 2013). There is evidence that

605 organic cows, required to obtain 30 percent of the daily matter intake (DMI) from grazing, are one third

- less likely to have ketosis than conventional animals (Hardeng & Edge, 2001). Grazing animals, both cows
- and sheep, also produce milk and meat that is higher in omega-3 fatty acids (Daley et al., 2010; Nuernberg
- et al., 2005; Wyss et al., 2010). There is evidence that omega-3 fatty acids improve energy metabolism
- 609 immediately after calving (Grossi et al., 2013), suggesting that animals who graze may be less likely to
- 610 succumb to ketosis.
- 611

Higher levels of neutral detergent fibers (the insoluble fibers in animal feed such as cellulose, 612 hemicellulose, and lignin) in feed are correlated with lower levels of serum NEFA (Litherland et al., 2013; 613 Van Soest et al., 1991). Lower levels of serum NEFA are negatively correlated with subclinical and clinical 614 615 ketosis in cows (Drackley & Cardoso, 2014; Duffield, 2000; Herdt, 2000; Vanholder et al., 2015). Litherland 616 et al. (2013) found that increased amounts of wheat straw in a pre- and postpartum diet in dairy cows 617 resulted in lower postpartum serum NEFA, suggesting healthier metabolism in postpartum cows. The 618 wheat straw helps to moderate the prepartum energy intake for animals. Animals overfed with energy 619 prepartum experienced a negative energy balance for longer into their lactation, which is the primary 620 driver of postpartum ketosis (Litherland et al., 2013). High-energy diets are typically low in both neutral 621 detergent fibers and acid detergent fibers, and are therefore nutrient dense (Agenäs et al., 2003; Mashek & Beede, 2000; Rabelo et al., 2003; Vandehaar et al., 1999). These high-energy diets lead to overeating, 622 623 providing significant energy before rumen fill. Drackley et al. (2014) demonstrated that cows fed high-624 energy diets during the dry period had greater serum concentrations of β -hydroxybutyrate, a ketone 625 related to ketosis. 626 627 Increasing forage and fibers in a ration leads to rumen fill and reduces DMI, including grain and 628 concentrates. There is evidence that feeding animals concentrates during the dry period does little more than needlessly fatten a cow (Grummer, 2008), leading to over-conditioned animals. Feeding concentrates 629 630 to dry cows in addition to silage exacerbates the negative energy balance after calving and elevates serum 631 concentrates of NEFA (Little et al., 2016), both of which correlate with incidence of postpartum ketosis. A 632 survey of organic and conventional farms in the United States showed that ketosis is less common on farms 633 where animals graze (Richert et al., 2013) and therefore achieve rumen fill through forage, lowering total 634 DMI in a ration. Drackley and Cardoso (2014) emphasized the need to formulate feed rations for dry cows to limit excess energy intake in the lead-up to calving. These new studies contradict the "steam-up" theory 635 of dry cow nutrition from the mid- and early twentieth century, which recommended increased levels of 636 637 grain in pre-transition cows (Boutflour, 1928; Grummer, 2008). 638 639 Finally, recent studies suggest that lower stocking densities, separate calving pens, and longer recovery 640 time for transition cows lowers rates of postpartum ketosis (Campler et al., 2019; Kaufman et al., 2016). 641 Providing transitioning cows with more space and longer recovery time allows animals to have longer lying periods, which increases rumination, promotes better feeding behavior, and reduces competition for 642 643 feed (Kaufman et al., 2016). Improved DMI and feeding post parturition leads to a shorter period of 644 negative energy balance and is associated with a lower incidence of ketosis (Campler et al., 2019). Campler et al. (2019) report that extended time in maternity pens reduces stress on animals following calving. 645

646

658 659

660

663 664

665

647 *Dehydration/electrolyte imbalance:*

648 Preventive measures that should be taken in order to avoid dehydration and electrolyte imbalance in

- 649 livestock include:
- -Proper nutrition and hydration of the animals, specifically the pregnant animals (particularly during the
 last third of gestation) and neonatal calves (Stoltenow and Vincent, 2003).

652 -Adequate environment for the mother and neonatal calves: avoiding overcrowding and contamination of

653 the space, maintaining proper sanitation dryness and cleanliness of the environment avoiding exposure to

- cold temperatures, rainfall and other stressful conditions (Stoltenow and Vincent, 2003).
- -Proper administration of colostrum with adequate content of immunoglobulin G (IgG) for the neonatal
- calves (<2 hours after birth); colostrum IgG concentration appears to be an important factor that affects
- 657 whether calves receive sufficient immunity from colostrum (Meganck et al., 2014).

Report Authorship

The following individuals were involved in research, data collection, writing, editing, and/or finalapproval of this report:

- Aura del Angel A Larson, Technical Coordinator, OMRI
 - Peter O. Bungum, Senior Technical Coordinator, OMRI

	Technical Evaluation Report	Glucose	Livestock
6	• Phoebe Judge, Technical Co	ordinator, OMRI	
7	•	or Bilingual Technical Coordinator, OMRI	
8	Amy Bradsher, Deputy Dire		
9	Doug Currier, Technical Dir		
0 1	Peggy Miars, Executive Dire		
2	1	ith Federal Acquisition Regulations (FAR) S	1 0
'3 '4	Personal Conflicts of Interest for Co	ntractor Employees Performing Acquisition	n Functions.
5		References	
6			
7		ns & Electrolytes "Plus" and Hydra-Lyte®.	
8 9	df and https://www.qcsup	n/media/product_attachments/attachmen ply.com/hydralyte-electrolyte-replacemen	
0	Accessed February 11, 2022		
1 2	Practice, 4(2), 233–251.	tosis in Dairy Cows. Veterinary Clinics of No	orth America: Food Animal
3	https://doi.org/10.1016/S0	749-0720(15)31046-X	
•	1 1	2021). Vita Jec K1. https://www.drugs.com/	/vet/vita-jec-k1.html.
	Accessed 2/15/2022.		
5	-	schall, E. F. (Eds. 3). (2012). Starch: chemistry	<i>and technology</i> . Academic
	Press.		
,	-	tate-of-the-art strategies and applied perspe	5
)	5	status and future trends. Critical Reviews in	1 Food Science and Nutrition,
)	60(12), 2052–2066.		
	https://doi.org/10.1080/10		
	of Glucose Production from	. G., Melero, J., & Moreno, J. (2020). Compar Maize Starch and Woody Biomass Residue	1
	Sciences, 10, 2946.		
	https://doi.org/10.3390/ap		
		tabolic needs for glucose and the role of glucose and	uconeogenesis. European
	Journal of Clinical Nutrition,		
	https://doi.org/10.1038/sj.		
	Da Silva, S., & Rodríguez, P	J. R., Benech, A., Acosta-Dibarrat, J., Martín. . (2015). Evaluation of three therapeutic alte	ernatives for the early
		cy toxaemia. Irish Veterinary Journal, 68(1), 2	25.
	https://doi.org/10.1186/s1		· · · · · · · · ·
		Jensen, M. B. (2019). The effect of transition	
		n dairy cows. <i>Journal of Dairy Science</i> , 102(8)	,/398-/40/.
	https://doi.org/10.3168/jd		
	-	& McArt, J. A. A. (2021). A randomized con	
	1 17 07	combination with dextrose as a treatment	tor hyperketonemia in dairy
	cows. Journal of Dairy Science		
	https://doi.org/10.3168/jd		
		v for the chemical and biological sciences. U	Iniversity Science Books.
		Diastereomers. Chemistry LibreTexts.	
		/Bookshelves/Organic_Chemistry/Organi	
		Tetrahedral_Centers/5.06%3A_Diastereom	
		rolyte therapy in ruminants. <i>Veterinary Clin</i>	ucs of North America: Food
	Animal Practice, 19(3), 557–5		
	https://doi.org/10.1016/S0		
	Coskun, O. (2016). Separation techn https://doi.org/10.14744/r	iques: Chromatography. <i>Northern Clinics of</i> aci.2016.32757	Istanbul, 3(2), 156–160.

719	Dagogo-Jack, S., & Alberti, K. G. M. M. (2002). Management of Diabetes Mellitus in Surgical Patients.
720	Diabetes Spectrum, 15(1), 44–48.
721	https://doi.org/10.2337/diaspect.15.1.44
722	Daley, C. A., Abbott, A., Doyle, P. S., Nader, G. A., & Larson, S. (2010). A review of fatty acid profiles and
723	antioxidant content in grass-fed and grain-fed beef. Nutrition Journal, 9(1), 10.
724	https://doi.org/10.1186/1475-2891-9-10
725	David Baird, G. (1982). Primary Ketosis in the High-Producing Dairy Cow: Clinical and Subclinical
726	Disorders, Treatment, Prevention, and Outlook. Journal of Dairy Science, 65(1), 1–10.
727	https://doi.org/10.3168/jds.S0022-0302(82)82146-2
728	de Souza, P. M., & Oliveira Magalhães, P. (2010). Application of microbial α-amylase in industry – A
729	review. Brazilian Journal of Microbiology: [Publication of the Brazilian Society for Microbiology], 41(4),
730	850-861.
731	https://doi.org/10.1590/S1517-83822010000400004
732	DHI Milieu Ltd., P. Ltd. (2008). Review of Annex IV of regulation (EC) No. 1907/2006 (Reach)- Evaluation
733	of Existing Entries in Annex IV.
734	https://ec.europa.eu/environment/chemicals/reach/pdf/6b_appendix_2.pdf
735	Dogru, Y. G. (2021). Is Low Dose Dextrose Prolotherapy as Effective as High Dose Dextrose Prolotherapy
736	in the Treatment of Lateral Epicondylitis?- A Double Blind- Ultrasound Guided- Randomized
737	Controlled Study (<i>Clinical Trial Registration No. NCT04680936</i>).
738	clinicaltrials.gov. https://clinicaltrials.gov/ct2/show/NCT04680936
739	Drackley, J. K., & Cardoso, F. C. (2014). Prepartum and postpartum nutritional management to optimize
740	fertility in high-yielding dairy cows in confined TMR systems. Animal, 8, 5–14.
741	https://doi.org/10.1017/S1751731114000731
742	Duffield, T. (2000). Subclinical Ketosis in Lactating Dairy Cattle. <i>Veterinary Clinics of North America: Food</i>
743	Animal Practice, 16(2), 231–253.
744	https://doi.org/10.1016/S0749-0720(15)30103-1
745	DuPont Industrial Biosciences. (2015, December 18). An Alpha-amylase EnzymePreparation Derived from
746	Bacillus licheniformis Expressing the Alpha-amylase Gene From Cytophaga sp. Is Generally
747	Recognized As Safe. https://www.fda.gov/media/97853/download
748	FDA. (2021). FDA U.S.A. Food & Drug Administration. Drugs@FDA: FDA-Approved Drugs. Retrieved
749	October 4, 2021 from US FDA:
750	https://www.accessdata.fda.gov/scripts/cder/daf/index.cfm?event=BasicSearch.process
751	Fellers, C. R. (1939). Dextrose in the Food Industries and Its Health Status. American Journal of Public Health
752	and the Nations Health, 29(2), 135–138.
753	https://doi.org/10.2105/AJPH.29.2.135
754	Ferraro, S. M., Mendoza, G. D., Miranda, L. A., & Gutiérrez, C. G. (2016). In vitro ruminal fermentation of
755	glycerol, propylene glycol and molasses combined with forages and their effect on glucose and
756	insulin blood plasma concentrations after an oral drench in sheep. Animal Feed Science and
757	<i>Technology</i> , 213, 74–80.
758	https://doi.org/10.1016/j.anifeedsci.2016.01.010
759	Ferreira, A. S., Santos, M. A. D., & Corrêa, G. F. (2013). Soil microbial response to glucose and phosphorus
760	addition under agricultural systems in the Brazilian Cerrado. Anais Da Academia Brasileira de
761	<i>Ciências</i> , 85(1), 395–403.
762	https://doi.org/10.1590/S0001-37652013005000021
763	Fisher Science Education. (2015). Glucose 15% Solution SDS. Retreived October 8, 2021 from:
764	https://betastatic.fishersci.com/content/dam/fishersci/en_US/documents/programs/education
765	/regulatory-documents/sds/chemicals/chemicals-g/S25338.pdf
766	Fujii, M., Murakami, Y., Karasawa, Y., Sumitomo, Y., Fujita, S., Koyama, M., Uda, S., Kubota, H., Inoue, H.,
767	Konishi, K., Oba, S., Ishii, S., & Kuroda, S. (2019). Logical design of oral glucose ingestion pattern
768	minimizing blood glucose in humans. Npj Systems Biology and Applications, 5(1), 1–11.
769	https://doi.org/10.1038/s41540-019-0108-1
770	Galant, A. L., Kaufman, R. C., & Wilson, J. D. (2015). Glucose: Detection and analysis. <i>Food Chemistry</i> , 188,
771	149–160.
772	https://doi.org/10.1016/j.foodchem.2015.04.071
· · -	$\mathbf{r} = \mathbf{r} + \mathbf{r}$

773	Gerloff, B. J. (2000). Dry Cow Management for the Prevention of Ketosis and Fatty Liver in Dairy Cows.
774	Veterinary Clinics of North America: Food Animal Practice, 16(2), 283–292.
775	https://doi.org/10.1016/S0749-0720(15)30106-7
776	Giri, B., Dey, S., Das, T., Sarkar, M., Banerjee, J., & Dash, S. K. (2018). Chronic hyperglycemia mediated
777	physiological alteration and metabolic distortion leads to organ dysfunction, infection, cancer
778	progression and other pathophysiological consequences: An update on glucose toxicity.
779	Biomedicine & Pharmacotherapy, 107, 306–328.
780	https://doi.org/10.1016/j.biopha.2018.07.157
781	Gocke, K., Mancera Pineda, J. E., & Vallejo, A. (2003). Heterotrophic microbial activity and organic matter
782	degradation in coastal lagoons of Colombia. <i>Revista de Biología Tropical</i> , 51(1), 85–98.
783	Gordon, J. L., LeBlanc, S. J., & Duffield, T. F. (2013). Ketosis Treatment in Lactating Dairy Cattle. Veterinary
784	Clinics of North America: Food Animal Practice, 29(2), 433–445.
785	https://doi.org/10.1016/j.cvfa.2013.03.001
786	Grossi, P., Bertoni, G., Cappelli, F. P., & Trevisi, E. (2013). Effects of the precalving administration of
787	omega-3 fatty acids alone or in combination with acetylsalicylic acid in periparturient dairy cows1.
788	Journal of Animal Science, 91(6), 2657–2666.
789	https://doi.org/10.2527/jas.2012-5661
790	Gunina, A., & Kuzyakov, Y. (2015). Sugars in soil and sweets for microorganisms: Review of origin,
791	content, composition and fate. <i>Soil Biology and Biochemistry</i> , 90, 87–100.
792	https://doi.org/10.1016/j.soilbio.2015.07.021
793 704	Hach Company. (2005). Glucose [Material Safety Data Sheet]. Retrieved September 12, 2021 from:
794 705	https://web.archive.org/web/20170101003021/http://www.chem.utoronto.ca/~pmeindl/labs/
795	msds%20files/glucose.pdf
796	Hardeng, F., & Edge, V. L. (2001). Mastitis, Ketosis, and Milk Fever in 31 Organic and 93 Conventional
797	Norwegian Dairy Herds. Journal of Dairy Science, 84(12), 2673–2679.
798	https://doi.org/10.3168/jds.50022-0302(01)74721-2
799	Hauser, R. A., Lackner, J. B., Steilen-Matias, D., & Harris, D. K. (2016). A Systematic Review of Dextrose
800	Prolotherapy for Chronic Musculoskeletal Pain. Clinical Medicine Insights: Arthritis and
801	Musculoskeletal Disorders, 9, CMAMD.S39160.
802	https://doi.org/10.4137/CMAMD.S39160
803	Havekes, C. D., Duffield, T. F., Carpenter, A. J., & DeVries, T. J. (2020). Effects of molasses-based liquid feed
804	supplementation to a high-straw dry cow diet on feed intake, health, and performance of dairy
805	cows across the transition period. <i>Journal of Dairy Science</i> , 103(6), 5070–5089.
806	https://doi.org/10.3168/jds.2019-18085
807	Herdt, T. H. (2000). Ruminant Adaptation to Negative Energy Balance. Veterinary Clinics of North America:
808	Food Animal Practice, 16(2), 215–230.
809	https://doi.org/10.1016/S0749-0720(15)30102-X
810	Herdt, T. H., & Emery, R. S. (1992). Therapy of Diseases of Ruminant Intermediary Metabolism. Veterinary
811	Clinics of North America: Food Animal Practice, 8(1), 91–106.
812	https://doi.org/10.1016/S0749-0720(15)30761-1
813	Hull, P. (2010). Glucose syrups: technology and applications. John Wiley & Sons.
814	https://doi.org/10.1002/9781444314748
815	International Starch Institute. (2021). International Starch: Production of corn starch. TM 18-5www - ISI
816	Technical Memorandum on Production of Corn Starch. Retreived September 17, 2021 from the
817	International Starch Institute:
818	http://www.starch.dk/isi/starch/tm18www-corn.htm
819	Jackson, E. B. (1995). Use of glucose syrups in the food industry. In M. W. Kearsley & S. Z. Dziedzic (Eds.),
820	Handbook of Starch Hydrolysis Products and their Derivatives (pp. 245–268). Springer US.
821	https://doi.org/10.1007/978-1-4615-2159-4_9
822	Kalyesubula, M., Rosov, A., Alon, T., Moallem, U., & Dvir, H. (2019). Intravenous Infusions of Glycerol
823	Versus Propylene Glycol for the Regulation of Negative Energy Balance in Sheep: A Randomized
824	Trial. Animals, 9(10), 731.
825	https://doi.org/10.3390/ani9100731

826 827 828	Kaufman, E. I., LeBlanc, S. J., McBride, B. W., Duffield, T. F., & DeVries, T. J. (2016). Association of rumination time with subclinical ketosis in transition dairy cows. <i>Journal of Dairy Science</i> , 99(7), 5604–5618.
829	https://doi.org/10.3168/jds.2015-10509
830 831	Kearsley, M. W., & Dziedzic, S. Z. (1995). Physical and chemical properties of glucose syrups. Handbook of Starch Hydrolysis Products and their Derivatives (pp. 129–154). Springer US.
832	https://doi.org/10.1007/978-1-4615-2159-4_5
833	Kirchman, D. L. (2003). The Contribution of Monomers and other Low-Molecular Weight Compounds to
834 835	the Flux of Dissolved Organic Material in Aquatic Ecosystems. <i>Aquatic Ecosystems</i> (pp. 217–241). Elsevier.
836	https://doi.org/10.1016/B978-012256371-3/50010-X
837	Kis, F., Maravić, N., Kertesz, S., & Šereš, Z. I. (2019). Life cycle assessment of liquid inverted sugar and
838	high-fructose corn syrup. Analecta Technica Szegedinensia, 13(1), 28–39.
839	https://doi.org/10.14232/analecta.2019.1.28-39
840 841	Klein, K. A., Clark, C., & Allen, A. L. (2002). Hypoglycemia in sick and moribund farmed elk calves. <i>The Canadian Veterinary Journal</i> , 43(10), 778–781.
842	Knapp, J. S., & Bromley-Challoner, K. C. A. (2003). 34–Recalcitrant organic compounds. D. Mara & N.
843	Horan (Eds.), Handbook of Water and Wastewater Microbiology (pp. 559-595). Academic Press.
844	https://doi.org/10.1016/B978-012470100-7/50035-2
845	Kupczyński, R., Szumny, A., Wujcikowska, K., & Pachura, N. (2020). Metabolism, Ketosis Treatment and
846	Milk Production after Using Glycerol in Dairy Cows: A Review. <i>Animals</i> , 10(8), 1379.
847	https://doi.org/10.3390/ani10081379
848	Lans, C., Turner, N., Khan, T., Brauer, G., & Boepple, W. (2007). Ethnoveterinary medicines used for
849 850	ruminants in British Columbia, Canada. <i>Journal of Ethnobiology and Ethnomedicine</i> , 3(1), 11. https://doi.org/10.1186/1746-4269-3-11
851	Litherland, N. B., da Silva, D. N. L., Hansen, W. P., Davis, L., Emanuele, S., & Blalock, H. (2013). Effects of
852	prepartum controlled-energy wheat straw and grass hay diets supplemented with starch or sugar
853	on periparturient dairy cow performance and lipid metabolism. Journal of Dairy Science, 96(5), 3050-
854	3063.
855	https://doi.org/10.3168/jds.2012-5998
856	Macrae, R., Robinson, R. K., & Sadler, M. J. (1993). Encyclopaedia of food science, food technology and nutrition.
857	Mann, S., Yepes, F. A. L., Behling-Kelly, E., & McArt, J. A. A. (2017). The effect of different treatments for
858	early-lactation hyperketonemia on blood β -hydroxybutyrate, plasma nonesterified fatty acids,
859	glucose, insulin, and glucagon in dairy cattle. <i>Journal of Dairy Science</i> , 100(8), 6470–6482.
860	https://doi.org/10.3168/jds.2016-12532
861	Meganck, V., Hoflack, G., & Opsomer, G. (2014). Advances in prevention and therapy of neonatal dairy calf
862	diarrhoea: a systematical review with emphasis on colostrum management and fluid therapy. Acta
863	Veterinaria Scandinavica, 56(1), 1-8.
864	McSherryt, B. J., Maplesdent, D. C., & Braniont, H. D. (1960). KETOSIS IN CATTLE-A REVIEW. The
865	Canadian Veterinary Journal, 1(5), 208.
866	Metzner, M., Hofmann, W., & Laiblin, C. (1993). Untersuchungen zur Wirksamkeit intravenös
867	verabreichter hoher Glukosemengen bei der Behandlung der Ketose des Rindes [The effectiveness
868	of intravenous administration of large quantities of glucose in the treatment of bovine ketosis].
869	Tierarztl Prax., 21(4):289-93.
870 871	Mironescu, M., & Mironescu, V. (2006). HYGIENIC ASPECTS AT THE PRODUCTION OF GLUCOSE SYRUPS THROUGH ACID HYDROLYSIS. FOOD TECHNOLOGY, 10.
872	Muralikrishna, G., & Nirmala, M. (2005). Cereal α-amylases – An overview. <i>Carbohydrate Polymers</i> , 60(2),
873	163-173.
874	https://doi.org/10.1016/j.carbpol.2004.12.002
875	Murphy, B., Horner, G., Tarcy, D., & Bylikin, S. (2014). Oxford IB Diploma Programme: Chemistry Course
876	Companion. Oxford University Press-Children.
877	NOP. (2000, December 21). National Organic Program; Amendments to the National List of Allowed and
878	Prohibited Substances (Livestock). Federal Register Number 65 FR 80547.
879	https://www.federalregister.gov/documents/2000/12/21/00-32257/national-organic-program

880	National Center of Biotechnology. (2021). D-Glucose. National Center of Biotechnology. Retreived September
881	9, 2021 from NCBI:
882	https://pubchem.ncbi.nlm.nih.gov/compound/D-Glucose.
883	Naylor, J. M. (1990). Oral Fluid Therapy in Neonatal Ruminants and Swine. Veterinary Clinics of North
884	America: Food Animal Practice, 6(1), 51–67.
885	https://doi.org/10.1016/S0749-0720(15)30894-X
886	Naylor, J. M. (1999). Oral Electrolyte Therapy. Veterinary Clinics of North America: Food Animal Practice, 15(3),
887	487-504. https://doi.org/10.1016/S0749-0720(15)30160-2
888	Nielsen, C. (2012). Amalyse modifications during production and application. <i>Technical University of</i>
889	Denmark.
890	Nuernberg, K., Nuernberg, G., Ender, K., Dannenberger, D., Schabbel, W., Grumbach, S., Zupp, W., &
891	Steinhart, H. (2005). Effect of grassvs. Concentrate feeding on the fatty acid profile of different fat
892	depots in lambs. European Journal of Lipid Science and Technology, 107(10), 737–745.
893	https://doi.org/10.1002/ejlt.200501141
894	Olempska-Beer, Z. (2004). ALPHA-AMYLASE FROM BACILLUS LICHENIFORMIS CONTAINING A
895	GENETICALLY ENGINEERED ALPHAAMYLASE GENE FROM B. LICHENIFORMIS
896	(THERMOSTABLE). Safety evaluation of certain food additives and contaminants, 3. Retreived
897	September 16, 2021 from FAO:
898	http://www.fao.org/fileadmin/templates/agns/pdf/jecfa/cta/61/alphaamylase.pdf
899	Olsen, H. S. (1995). Enzymatic production of glucose syrups. In M. W. Kearsley & S. Z. Dziedzic (Eds.),
900	Handbook of Starch Hydrolysis Products and their Derivatives (pp. 26–64). Springer US.
901	https://doi.org/10.1007/978-1-4615-2159-4_2
902	Padmanabhan, P., Padmanabhan, S., DeRito, C., Gray, A., Gannon, D., Snape, J. R., Tsai, C. S., Park,
903	W., Jeon, C., & Madsen, E. L. (2003). Respiration of 13 C-Labeled Substrates Added to Soil in the
904	Field and Subsequent 16S rRNA Gene Analysis of 13 C-Labeled Soil DNA. Applied and
905	Environmental Microbiology, 69(3), 1614–1622.
906	https://doi.org/10.1128/AEM.69.3.1614-1622.2003
907	Organic Materials Review Institute (OMRI). (2021). Internal analysis of livestock healthcare products
908	ingredients. Eugene, Oregon: unpublished.
909	Parajuli, B. R., & Sanjib, K. (2021). A concept of fusion medicine-where Ayurveda meets Allopathy. <i>Journal</i>
910	of Karnali Academy of Health Sciences. 4 (2).
911	Piantoni, P., & Allen, M. S. (2015). Evaluation of propylene glycol and glycerol infusions as treatments for
912	ketosis in dairy cows. Journal of Dairy Science, 98(8), 5429–5439.
913	https://doi.org/10.3168/jds.2015-9476
914	Precision Feed Technologies, LLC (2021). <i>Products: Gut Candy</i> . https://www.gutcandy.com/products.html.
915	Accessed 2/15/2022.
916	Pronk, M. I., & Leclercq, C. (2004). A-AMYLASE FROM BACILLUS LICHENIFORMIS CONTAINING A
917	GENETICALLY ENGINEERED a-AMYLASE GENE FROM B. LICHENIFORMIS. World Health
918	Organization, Geneva, 2004 IPCS – International Programme on Chemical Safety.
919	Purolite. (2021). Chromatographic Separation Process. Retrieved September 29, 2021 from Purolite:
920	http://www.purolite.com/index/core-technologies/industry/food-and-beverage/sweetener-
921	applications/corn-sweetener-refining-with-ion-exchange-resins/color-taste-and-odor-
922	removal/chromatographic-separation-process
923	Reddy, N. S., Nimmagadda, A., & Rao, K. (2003). An overview of the microbial α-amylase family.
924	2(12), African journal of biotechnology, 2(12), 645-648.
925	Reid, S. R., & Losek, J. D. (2009). Role for Early Use of Intravenous Dextrose. <i>Pediatric Emergency Care</i> , 25(1),
926	7.
927	Reinhardt, C., & Faris, B. (2014). NUTRITION OF MEAT ANIMALS Ruminants. In M. Dikeman & C.
928	Devine (Eds.), Encyclopedia of Meat Sciences (Second Edition) (pp. 471–479). Academic Press.
929	https://doi.org/10.1016/B978-0-12-384731-7.00022-2
930	Renouf, M. A., Wegener, M. K., & Nielsen, L. K. (2008). An environmental life cycle assessment comparing
931	Australian sugarcane with US corn and UK sugar beet as producers of sugars for fermentation.
932	Biomass and Bioenergy, 32(12), 1144–1155.
933	https://doi.org/10.1016/j.biombioe.2008.02.012

934 935	Richert, R. M., Cicconi, K. M., Gamroth, M. J., Schukken, Y. H., Stiglbauer, K. E., & Ruegg, P. L. (2013). Risk factors for clinical mastitis, ketosis, and pneumonia in dairy cattle on organic and small
936	conventional farms in the United States. Journal of Dairy Science, 96(7), 4269-4285.
937	https://doi.org/10.3168/jds.2012-5980
938	Rozance, P. J., & Hay, W. W., Jr (2010). Describing hypoglycemiadefinition or operational
939	threshold?. Early human development, 86(5), 275–280.
940	https://doi.org/10.1016/j.earlhumdev.2010.05.002
941	Sargison, N. D. (2007). "Pregnancy Toxaemia". Diseases of sheep, 7, 359-362.
942	Schenck, F. W. (2000). Glucose and glucose-containing syrups. Ullmann's Encyclopedia of industrial
943	Chemistry.
944	Shahbandeh, M. (2021). Corn in the U.S. Statistics & Facts. Retreived September 29, 2021 from Statista:
945	https://www.statista.com/topics/986/corn/
946	Silano, V., Bolognesi, C., Castle, L., Chipman, K., Cravedi, J., Fowler, P., Franz, R., Grob, K., Gürtler, R.,
947	Husøy, T., Kärenlampi, S., Mennes, W., Milana, M. R., Pfaff, K., Riviere, G., Srinivasan, J., Tavares
948	Poças, M. de F., Tlustos, C., Wölfle, D., Engel, K. (2018). Safety evaluation of the food enzyme a-
949 950	amylase from a genetically modified Aspergillus niger (strain NZYM-SB). <i>EFSA Journal,</i> 16(7), e05320.
951	https://doi.org/10.2903/j.efsa.2018.5320
952	Skoog, A., Biddanda, B., & Benner, R. (1999). Bacterial Utilization of Dissolved Glucose in the Upper Water
953	Column of the Gulf of Mexico. <i>Limnology and Oceanography</i> , 44(7), 1625–1633.
954	https://doi.org/10.4319/lo.1999.44.7.1625
955	Smith, G. W. (2009). Treatment of Calf Diarrhea: Oral Fluid Therapy. Veterinary Clinics of North America:
956	Food Animal Practice, 25(1), 55–72.
957	https://doi.org/10.1016/j.cvfa.2008.10.006
958	Stock Show Secrets (2022). Attitude Adjustment Paste for Livestock.
959	https://www.valleyvet.com/library/lib_45710Attitudeadjustment.pdf. Accessed 2/15/2022.
960	Stoltenow, C. L., & Vincent, L. L. (2003). Calf scours: causes, prevention, treatment.
961	Srivastava, H. K., Wolfgang, F., & Rodriguez, J. D. (2016). Expanding the analytical toolbox for identity
962	testing of pharmaceutical ingredients: Spectroscopic screening of dextrose using portable Raman
963	and near infrared spectrometers. <i>Analytica chimica acta</i> , 914, 91-99.
964	https://doi.org/10.1016/j.aca.2016.01.061
965	University of Reading NCBE. (2018). <i>Alpha-amylase (Termamyl)</i> . <i>ENZYMES FOR EDUCATION - a-Amylase</i> .
966	Retreived: October 6, 2021 from NCBE:
967 968	http://www.ncbe.reading.ac.uk/materials/enzymes/termamyl.html US EPA, O. (2004). U.S. Environmental Protection Agency (EPA) (2004). "List 4A Minimal Risk Inert
908 969	Ingredients – By Chemical Name Updated August 2004". Retreived September 9, 2021 from US
909 970	Environmental Protection Agency:
970 971	https://www.epa.gov/sites/default/files/2015-10/documents/inerts_list4aname.pdf
972	US EPA, O. (2020, March 26). <i>Commodity Inert Ingredients [Data and Tools]</i> . Retreived September 9, 2021 from
973	US Environmental Protection Agency:
974	https://www.epa.gov/pesticide-registration/commodity-inert-ingredients
975	USDA. (2021). Technical evaluation report: Propylene Glycol. Washington, D.C.: National Organic Program.
976	USDA-Economic Research Service. (2020). <i>Table 37- U.S. dextrose supply and use, by calendar year</i> . Retreived:
977	September 29, 2021 from US Department of Agriculture, Economic Research Service:
978	https://www.ers.usda.gov/data-products/sugar-and-sweeteners-yearbook-tables/sugar-and-
979	sweeteners-yearbook-
980	tables/#World%20and%20U.S.%20Sugar%20and%20Corn%20Sweetener%20Prices
981	Van Soest, P. J., Robertson, J. B., & Lewis, B. A. (1991). Methods for Dietary Fiber, Neutral Detergent Fiber,
982	and Nonstarch Polysaccharides in Relation to Animal Nutrition. Journal of Dairy Science, 74(10),
983	3583-3597.
984	https://doi.org/10.3168/jds.S0022-0302(91)78551-2
985	Vanholder, T., Papen, J., Bemers, R., Vertenten, G., & Berge, A. C. B. (2015). Risk factors for subclinical and
986	clinical ketosis and association with production parameters in dairy cows in the Netherlands.
987	Journal of Dairy Science, 98(2), 880–888.
988	https://doi.org/10.3168/jds.2014-8362

989	Vercalsteren, A., Dils, E., & Boonen, K. (2012). Life Cycle Assessment study of starch products for the
990	European starch industry association (AAF): Sector study. Flemish Institute for Technological Research
991	NV, Boeretang.
992	VetOne®. 2022. Dextrose 50% Injection.
993	http://www.vetone.net/Default/GetFile?id=3f884329-1414-e411-b319-0024e878511a. Accessed
994	February 11, 2022.
995	Vickers, L. A., Weary, D. M., Veira, D. M., & von Keyserlingk, M. A. G. (2013). Feeding a higher forage diet
996	prepartum decreases incidences of subclinical ketosis in transition dairy cows1. Journal of Animal
997	Science, 91(2), 886–894.
998	https://doi.org/10.2527/jas.2011-4349
999	Wagner, S. A., & Schimek, D. E. (2010). Evaluation of the effect of bolus administration of 50% dextrose
1000	solution on measures of electrolyte and energy balance in postpartum dairy cows. American Journal
1001	of Veterinary Research, 71(9), 1074–1080. https://doi.org/10.2460/ajvr.71.9.1074
1002	Walter, G. (2020). Colisepticemia - Generalized Conditions. Merck Veterinary Manual.
1003	https://www.merckvetmanual.com/generalized-
1004	conditions/colisepticemia/colisepticemia?query=hypoglycemia%20treatment
1005	Wyss, U., Munger, A., & Collomb, M. (2010). Variation of Fatty Acid Content in Grass and Milk During the
1006	Grazing Season. Grassland Science in Europe, 15, 422-424.
1007	Zainab, A., Modu, S., & Falmata, A. S. (2011). Laboratory scale production of glucose syrup by the
1008	enzymatic hydrolysis of starch made from maize, millet and sorghum. <i>Biokemistri</i> , 23(1).
1009	Zhang, G., & Ametaj, B. N. (2020). Ketosis an Old Story Under a New Approach. Dairy, 1(1), 42–60.
1010	https://doi.org/10.3390/dairy1010005
1011	Zhang, R., Ma, S., Li, L., Zhang, M., Tian, S., Wang, D., Liu, K., Liu, H., Zhu, W., & Wang, X. (2021).
1012	Comprehensive utilization of corn starch processing by-products: A review. Grain & Oil Science and
1013	Technology.
1014	https://doi.org/10.1016/j.gaost.2021.08.003
1015	
1016	