Parasiticides: Fenbendazole, Ivermectin, Moxidectin
Livestock

Identification of Petitioned Substance*

Chemical Names:


Fenbendazole: methyl N-(6-phenylsulfanyl-1H-benzimidazol-2-yl) carbamate

Ivermectin: 22,23-dihydroavermectin B1a + 22,23-dihydroavermectin B1b

Thiabendazole: 4-(1H-1,3-benzodiazol-2-yl)-1,3-thiazole

Albendazole: Methyl [5-(propylthio)-1H-benzoimidazol-2-yl]carbamate

Levamisole: (S)-6-Phenyl-2,3,5,6-tetrahydroimidazo[2,1-b][1,3]thiazole

Morantel tartrate: 2,3-dihydroxybutanedioic acid; 1-methyl-2-[(E)-2-(3-methylthiophen-2-yl)ethenyl]-5,6-dihydro-4H-pyrimidine

Pyrantel: 4-(3-carboxy-2-hydroxynaphthalen-1-methyl)-3-hydroxynaphthalene-2-carboxylic acid; 1-methyl-2-[(E)-2-thiophen-2-ylethenyl]-5,6-dihydro-4H-pyrimidine

Doramectin: 1,25-cyclohexyl-5-O-demethyl-25-de(1-methylpropyl)avermectin A1a

Eprinomectin: (4'R)-4''-(Acetylamino)-4''-deoxyavermectin B1

Piperazine: Hexahydropyrazine; Piperazidine; Diethylenediamine

Other Name:

Moxidectin: Milbemycin B
Fenbendazole
Ivermectin: Dihydroavermectin

Trade Names:

Moxidectin: Equest, Cydectin, ProHeart 6
Fenbendazole: Panacur, Safe Guard

Ivermectin: Heart Guard, Sklice, Stomectol, Ivomec, Mectizan, Ivexterm, Scabo 6
Thiabendazole: Mintezol, Tresaderm, Arbotect Albendazole: Albenza

CAS Numbers:

Moxidectin: 113507-06-5;
Fenbendazole: 43210-67-9;
Ivermectin: 70288-86-7
Thiabendazole: 148-79-8 Albendazole: 54965-21-8
Levamisole: 14769-72-4 Morantel tartrate: 26155-31-7
Pyrantel: 22204-24-6 Doramectin: 117704-25-3
Eprinomectin: 123997-26-2 Piperazine: 110-85-0

Other Codes:

Moxidectin: Pubchem CID 16760141; InChI Key: YZBLFMPOMVTDJY-CBYMMZEQSA-N; ChemSpider 167363424
Fenbendazole: PubChem CID 3334; InChI Key HDDSHPAOJDJKP-UHFFFAOYSA-N; ChemSpider: 3217
Ivermectin: PubChem CID 4330618; InChI Key A5ZSNMRSAGSSBSNP-UHFFFAOYSA-N; ChemSpider 7988461
Thiabendazole: PubChem: CID 5430
Albendazole: PubChem: CID 2082
Levamisole: PubChem: CID 26879
Morantel tartrate: PubChem: CID 6419965
Pyrantel: PubChem: CID 5281033
Doramectin: PubChem: CID 9832750
Eprinomectin: PubChem: CID 6426924
Piperazine: PubChem: CID 4837

*Substances within the scope of this review are in bold
The Organic Foods Production Act (OFPA), 7 U.S.C. 6501 et seq., authorizes the establishment of the National List of allowed and prohibited substances. Exemptions and prohibitions granted under the OFPA are required to be reviewed every 5 years by the National Organic Standards Board (NOSB). The NOSB requested a Technical Advisory Panel (TAP) review of parasiticides in 1995 (NOP, 1995). At the time, ivermectin, fenbendazole and levamisole were under consideration by the NOSB for addition to the National List, § 205.603 Synthetic substances allowed for use in organic livestock production.

The National Organic Standards Board (NOSB) considered the use of parasiticides during its February, 1999 meeting (NOP, 1999a). A TAP review was accepted by the NOSB for parasiticides, November 25, 1999 (NOP, 1999b). Levamisole was reviewed by the NOSB in 1999, but failed to obtain the NOSB’s recommendation and was subsequently prohibited. Ivermectin was the first parasiticide included in the National List of Allowed and Prohibited Substances by the same final rule establishing the National Organic Program (NOP, 2000). It was listed as follows:

§ 205.603 Synthetic substances allowed for use in organic livestock production as

(a) medical treatment

(12) parasiticides—ivermectin—prohibited in slaughter stock, allowed in emergency treatment for dairy and breeder stock when organic system plan-approved preventive management does not prevent infestation. Milk or milk products from a treated animal cannot be labeled as provided for in subpart D of this part for 90 days following treatment. In breeder stock, treatment cannot occur during the last third of gestation if the progeny will be sold as organic and must not be used during the lactation period of breeding stock.

In a subsequent proposed rule, a petition for a second parasiticide, moxidectin, as a medical treatment for use in organic livestock production to control internal and external parasites was considered by the NOSB. The NOSB recommendation for adding moxidectin to the National List and a ruling by the US Agricultural Secretary preventing adoption of this recommendation were also published (NOP, 2006; NOP, 2003). Although the NOSB approved addition of moxidectin to the National List, the US Agriculture Secretary could not accept NOSB’s recommendation because moxidectin was labeled as a macrolide antibiotic (§205.238(c)(1), §205.238(c)(7), 7 USC Sec. 6517).

In a final rule, the exemption for ivermectin was renewed on October 21, 2007 (NOP, 2007a).

The exclusion of moxidectin was addressed in a final rule amending the National List (NOP, 2007b). Moxidectin and its precursor nemadectin are members of a group of compounds called macrolides. Macrolides contain a signature molecular structure called a macrolide lactone ring. Based on their molecular characteristics, macrolides are divided into two chemical groups, the erythromycins and the polyenes. Moxidectin and nemadectin are members of the polyene group of chemical products. The polyenes unlike their erythromycin counterparts do not possess antibiotic properties. They are inactive against bacteria and not considered antibiotics sensu stricto an antibiotic is a type of antimicrobial substance used specifically against bacteria (Hamilton-Miller, 1973). As a result of comments received by the NOP, proposed rulemaking was initiated to authorize moxidectin as a livestock medication to control internal parasites (NOP, 2007b).

A petition for inclusion of fenbendazole on the National List was received by the NOP, March 23, 2007 (NOP, 2007c). Subsequently, a proposed rule addressed NOSB recommendations to establish exemptions (uses) for two substances, fenbendazole and moxidectin, on the National List as parasiticides in organic livestock production (NOP, 2011). A final rule established practice for the use of parasiticides and exemptions (uses) for fenbendazole and moxidectin (NOP, 2012):


§ 205.238 Livestock health care practice standard.

(b) Parasiticides allowed under § 205.603 may be used on:
(1) Breeder stock, when used prior to the last third of gestation but not during lactation for progeny that are to be sold, labeled, or represented as organically produced; and

(2) Dairy stock, when used a minimum of 90 days prior to the production of milk or milk products that are to be sold, labeled, or represented as organic.

(c) The producer of an organic livestock operation must not:

(4) Administer synthetic parasiticides on a routine basis;

(5) Administer synthetic parasiticides to slaughter stock;

(7) Withhold medical treatment from a sick animal in an effort to preserve its organic status. All appropriate medications must be used to restore an animal to health when methods acceptable to organic production fail. Livestock treated with a prohibited substance must be clearly identified and shall not be sold, labeled, or represented as organically produced.

§ 205.603 Synthetic substances allowed for use in organic livestock production.

(a) As medical treatments as applicable.

(18) Parasiticides—Prohibited in slaughter stock, allowed in emergency treatment for dairy and breeder stock when organic system plan-approved preventive management does not prevent infestation. Milk or milk products from a treated animal cannot be labeled as provided for in subpart D of this part for 90 days following treatment. In breeder stock, treatment cannot occur during the last third of gestation if the progeny will be sold as organic and must not be used during the lactation period for breeding stock.

(i) Fenbendazole (CAS # 43210–67–9)—only for use by or on the lawful written order of a licensed veterinarian.

(ii) Ivermectin (CAS # 70288–86–7).

(iii) Moxidectin (CAS # 113507–06–5)—for control of internal parasites only.

Characterization of Petitioned Substance

In veterinary medicine the term parasiticide refers to anthelmintic drugs, although ivermectin and moxidectin are also effective against arthropod parasites. Anthelmintics are medications capable of causing the evacuation of parasitic intestinal worms. Fenbendazole, ivermectin and moxidectin are the only anthelmintics approved for use in organic livestock production. They represent two of five anthelmintic drug classes differentiated by their chemical structures. The five known classes of livestock anthelmintics are benzimidazoles, imidazothiazoles, tetrahydropyrimidines, macrocyclic lactones and piperazines (Table 1). Each drug targets a vital system of the parasitic worm to cause incapacitation, death and excretion.

Including fenbendazole, ivermectin and moxidectin, there are eleven parasiticides currently approved by the US Food and Drug Administration Center for Veterinary Medicine for use in livestock production. All available parasiticides including fenbendazole, moxidectin and ivermectin are subject to parasiticide resistance. Populations of naturally drug resistant worms and their eggs present in dairies, stockyards, barns, forages, fields and in infested livestock can cause the failure of anthelmintics to effectively remove parasites from infested animals. If one drug is shown to be ineffective because of resistance, producers and veterinarians can chose a different drug that is likely to be effective (Martin, 1997). Organic livestock production does not require the use of parasiticides; however, information on eight additional parasiticides is included to provide context for the “emergency toolkit” of parasiticides available to livestock producers for chemically controlling parasitic nematodes (Table 1).

Fenbendazole is the only benzimidazole approved for use in organic livestock production. Two other benzimidazoles approved by the US Food and Drug Administration are thia bendazole and albendazole. Thia bendazole was the first to be described in 1961. It was selected from several hundred analogous compounds with broad spectrum anthelmintic and larvocidal activity (Fig 1). Its potency coupled with the absence of activity toward other microorganisms and negligible mammalian toxicity provided a basis for using this compound...
commercially. The mode of action of thiabendazole was not understood at the time of its discovery (Brown et al., 1961).

Table 1 Anthelmintics approved in the United States for Livestock*

<table>
<thead>
<tr>
<th>Group</th>
<th>Active Ingredient</th>
<th>Manufacturer(s)-Trade Name***</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Benzimidazoles</strong></td>
<td>Thiabendazole</td>
<td>Merial Ltd.-Thiabendazole Sheep &amp; Goat Wormer, Thiazenzole, Omnizole, TBZ Cattle Wormer Thibenzoze; ADM Alliance Nutrition, Inc.-E-Z-X-Wormer</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Albenzole, Zoeitis-Valbazeen</td>
</tr>
<tr>
<td><strong>Fenbendazole</strong></td>
<td></td>
<td>Intervet (Merck)-Panacur®, Safe-Guard®, Lincomix; Virbac-Purina Worm-A-Rest Litter Pack; Zoetis-BMD®/Safe-Guard®</td>
</tr>
<tr>
<td><strong>Imidazothiazoles</strong></td>
<td>Levamisole</td>
<td>Zoetis-Riperacol, Tramisol; Intervet (Merck)-Levasole, Tramisol; Agri Laboratories-Prohibit, levamisole phosphate; Cross Vetpharm Group, Ltd.-Levamisole hydrochloride</td>
</tr>
<tr>
<td><strong>Tetrahydropyrimidines</strong></td>
<td>Morantel tartrate</td>
<td>Phibro Animal Health Corp.-Rumatel; Zoetis, Inc.-Rumatel, Paratect Flex</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Pyrantel</td>
</tr>
<tr>
<td><strong>Macrocyclic lactones</strong></td>
<td>Ivermectin</td>
<td>Merial Ltd.-Ivomec .27% Injection Grower And Feeder Pigs; Bayer HealthCare LLC, Animal Health Division-Phoeinctin™; Norbrook Laboratories Ltd-Noromectin Pour-On for Cattle; Cross Vetpharm Group Ltd.-Bimectin Pour-On; First Priority, Inc.-Primectin™ Drench for Sheep; Privermectin; SmartVet USA, Inc., Ecomectin; Norbrook Laboratories Ltd.-Noromectin; Sparhawk Laboratories, Inc.-SparMectin Plus Clorsulon</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Doramectin</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Eprinomectin</td>
</tr>
<tr>
<td><strong>Piperazines</strong></td>
<td>Piperazine</td>
<td>Fleming Laboratories-Pig Wormer, Wazine</td>
</tr>
</tbody>
</table>

Broader spectrum carbamate sulphides and sulphoxide benzimidazoles, respectively fenbendazole and albendazole with high efficacy against lungworms and larvical inhibition of *Ostertagia ostertagi* were introduced in the mid-1970s (Table 1; Fig 2.). More effective and marketable benzimidazoles have not been found (McKellar and Scott, 1990).

![Fig 1 Thiabendazole](Brown et al., 1961)

Levamisole is not currently approved for use in organic livestock production. It is also known as tetramisole, a derivative of 6-arylimidazo[2,1-b]thiazole, and the only member of the imidazothiazole class of anthelmintics.
approved by the FDA and marketed in the United States. The result of screening a large series of compounds, levamisole is active against parasites of sheep and chickens (Raeymakers et al., 1966; Merck, 1983; Table 1; Fig. 3).

Fenbendazole

Albendazole

Fig 2. Carbamate sulphide and sulphone benzimidazoles (McKellar and Scott, 1990)

Neither pyrantel tartrate nor morantel are approved for use in organic livestock production. Both are members of the tetrahydropyrimidine class of parasiticides. Morantel is the methyl derivative of pyrantel (Bogan and Armour, 1980). The efficacy of pyrantel as a veterinary anthelmintic was first described in 1966, shortly after the introduction of levamisole. Pyrantel is an imidazothiazole-derived tetrahydropyrimidine with a broad spectrum of activity against immature and adult nematodes (Fig 4; Kopp et al., 2008). Morantel is more potent than pyrantel and requires a lower dose rate for its anthelmintic effect. It is generally formulated as a tartrate salt (Table 1; Fig 4; Lanusse and Pritchard, 1993).

The avermectins and milbemycins are anthelmintic macrocyclic lactones derived from the Streptomycetaceae family of Actinobacteria (Prichard et al., 2012; Hamilton-Miller, 1973). They are members of the polyene family of antimicrobial substances (Hamilton-Miller, 1973). Four veterinary drugs in this class are approved for use by the FDA: ivermectin, doramectin, eprinomectin and moxidectin (Table 1; Fig 5).

Ivermectin is approved for use in organic livestock production. Ivermectin was the first of the macrocyclic lactone anthelmintics to be discovered. It is a semi-synthetic chemically reduced 22,23-dihydro derivative of abamectin (Campbell et al., 1983). Abamectin is produced by fermentation of the actinomycete, Streptomyces avermilibis which was first isolated from soil in Japan. Abamectin is a mixture of avermectin B$_{1a}$ and avermectin B$_{1b}$ (Stapley, E.O. and Woodruff, H.B., 1982, Prichard et al., 2012). Doramectin was initially isolated through a process called “mutational biosynthesis.” Briefly, mutant strains of Streptomyces avermilibis lacking branched chain 2-oxo-acid dehydrogenase activity were isolated, cultured and provided with an alternative carboxylic acid as a nutrient source. Fractions of broth from cultures of these strains were then tested for anthelmintic activity.
One fraction contained Doramectin—25-cyclohexyl-5-O-demethyl-25-de(1-methylpropyl) avermectin A1a (Goudie et al., 1993; Dutton et al., 1990). An increased frequency in homologous DNA recombination and relaxation of double stranded DNA repair in stationary phase bacteria under nutritional stress is thought to be the mechanism for mutational biosynthesis (Aravind and Koonin, 2000, Lopez-Olmos et al., 2012). Doramectin is not approved for use in organic livestock production.

![Fig 5. Avermectins and Miblemycin](image)

**Table 2 Physical and Chemical Properties of the Veterinary Parasiticides**

<table>
<thead>
<tr>
<th>Drug</th>
<th>Formula</th>
<th>Mol. Wt. (grams/mole)</th>
<th>Melting/Boiling Point, °C</th>
<th>Appearance</th>
<th>Solubility</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thiabendazole&lt;sup&gt;1&lt;/sup&gt;</td>
<td>C&lt;sub&gt;10&lt;/sub&gt;H&lt;sub&gt;10&lt;/sub&gt;N&lt;sub&gt;3&lt;/sub&gt;S</td>
<td>201.25</td>
<td>304-305</td>
<td>White to tan crystals</td>
<td>50 mg/L @25°C in water</td>
</tr>
<tr>
<td>Albendazole&lt;sup&gt;2&lt;/sup&gt;</td>
<td>C&lt;sub&gt;12&lt;/sub&gt;H&lt;sub&gt;15&lt;/sub&gt;N&lt;sub&gt;3&lt;/sub&gt;O&lt;sub&gt;2&lt;/sub&gt;S</td>
<td>265.33</td>
<td>208-210</td>
<td>Colorless crystals</td>
<td>41 mg/L @25°C in water</td>
</tr>
<tr>
<td>Fenbendazole&lt;sup&gt;3&lt;/sup&gt;</td>
<td>C&lt;sub&gt;15&lt;/sub&gt;H&lt;sub&gt;13&lt;/sub&gt;N&lt;sub&gt;3&lt;/sub&gt;O&lt;sub&gt;2&lt;/sub&gt;S</td>
<td>299.35</td>
<td>233</td>
<td>White to tan powder</td>
<td>Insoluble in water</td>
</tr>
<tr>
<td>Levamisole&lt;sup&gt;4&lt;/sup&gt;</td>
<td>C&lt;sub&gt;11&lt;/sub&gt;H&lt;sub&gt;12&lt;/sub&gt;N&lt;sub&gt;2&lt;/sub&gt;S</td>
<td>204.29</td>
<td>227-227.5</td>
<td>White to tan powder</td>
<td>210 mg/mL in water</td>
</tr>
<tr>
<td>Morantel tartrate&lt;sup&gt;5&lt;/sup&gt;</td>
<td>C&lt;sub&gt;16&lt;/sub&gt;H&lt;sub&gt;22&lt;/sub&gt;N&lt;sub&gt;2&lt;/sub&gt;O&lt;sub&gt;6&lt;/sub&gt;S</td>
<td>370.42</td>
<td>167-172</td>
<td>White or pale yellow crystalline powder</td>
<td>Very soluble in water</td>
</tr>
<tr>
<td>Pyrantel&lt;sup&gt;6&lt;/sup&gt;</td>
<td>C&lt;sub&gt;11&lt;/sub&gt;H&lt;sub&gt;14&lt;/sub&gt;N&lt;sub&gt;2&lt;/sub&gt;S</td>
<td>206.31</td>
<td>178-179</td>
<td>Yellow crystals</td>
<td>Insoluble in water</td>
</tr>
<tr>
<td>Ivermectin&lt;sup&gt;7&lt;/sup&gt;</td>
<td>C&lt;sub&gt;48&lt;/sub&gt;H&lt;sub&gt;74&lt;/sub&gt;O&lt;sub&gt;14&lt;/sub&gt;</td>
<td>875.10</td>
<td>155</td>
<td>Off white powder</td>
<td>Insoluble in water, soluble in methanol or ethanol</td>
</tr>
<tr>
<td>Doramectin&lt;sup&gt;8&lt;/sup&gt;</td>
<td>C&lt;sub&gt;48&lt;/sub&gt;H&lt;sub&gt;74&lt;/sub&gt;O&lt;sub&gt;14&lt;/sub&gt;</td>
<td>899.14</td>
<td>160.5-162.2</td>
<td>White to tan powder</td>
<td>0.003 g/L @25°C in water, very low solubility in water</td>
</tr>
<tr>
<td>Eprinomectin&lt;sup&gt;9&lt;/sup&gt;</td>
<td>C&lt;sub&gt;48&lt;/sub&gt;H&lt;sub&gt;74&lt;/sub&gt;O&lt;sub&gt;14&lt;/sub&gt;</td>
<td>914.14</td>
<td>173</td>
<td>White crystalline solid</td>
<td>0.0035 g/L @25oC in water, very low solubility in water</td>
</tr>
<tr>
<td>Moxidectin&lt;sup&gt;10&lt;/sup&gt;</td>
<td>C&lt;sub&gt;37&lt;/sub&gt;H&lt;sub&gt;53&lt;/sub&gt;NO&lt;sub&gt;8&lt;/sub&gt;</td>
<td>639.84</td>
<td>145-154</td>
<td>White to pale yellow crystalline powder</td>
<td>0.51 mg/L in water</td>
</tr>
<tr>
<td>Piperazine&lt;sup&gt;11&lt;/sup&gt;</td>
<td>C&lt;sub&gt;4&lt;/sub&gt;H&lt;sub&gt;10&lt;/sub&gt;N&lt;sub&gt;2&lt;/sub&gt;</td>
<td>86.14</td>
<td>106/146</td>
<td>Leaflets from alcohol</td>
<td>Soluble in water</td>
</tr>
</tbody>
</table>


Eprinomectin is not approved for use in organic livestock production, but was developed in an effort to find a safe and efficacious anthelmintic macrolide for use in dairy production. A large number of synthetic ivermectin
analogs were screened to identify eprinomectin, 4''-epi-acetylamino-4''-deoxy-avermectin B1. It was chosen for its wide therapeutic index and lowest residue level in milk (Shoop et al., 1996).

Moxidectin is the only milbemycin approved for use in organic livestock production (Takiguchi et al., 1980). Moxidectin, a derivative of nemadectin is a chemically modified Streptomyces cyanogriseus fermentation product (Asato and France, 1990). Moxidectin is related to ivermectin, but lacks a disaccharide moiety and has an O-methyl substituent at the 23-position (Deng et al., 1991).

![Fig. 6 Piperazine](image)

Piperazine is not currently approved for use in organic livestock production. It is prepared by the action of alcoholic ammonia on ethylene chloride, the reduction of parazine with sodium alcohol and the catalytic deamination of diethylenetriamine and ethylene diamine (Fig 6; Merck, 1983). Piperazine dihydrochloride, piperazine sulfate and piperazine phosphate are effective anthelmintics when used as feed additives in hogs (Guthrie and Briggs, 1956; Praslicka et al., 1997; Steffan et al., 1988).

**Source or Origin of the Substance:**

As veterinary drugs, parasiticides are articles intended for use in treatment or prevention of disease in animals (Section 201(g)(1)(B) & (C) of the Federal Food, Drug, and Cosmetic Act [21 U.S.C. 321(g)(1)(B) & (C)]). The Federal Food, Drug and Cosmetic Act gives the US Food and Drug Administration (FDA) legal authority to approve and regulate veterinary drugs for animals. FDA’s Center for Veterinary Medicine (CVM) approves and regulates all new animal drugs. An approved animal drug is one that has gone through the FDA’s new animal drug application (NADA) process and has been stamped approved by the CVM. CVM’s approval means that the drug is safe and effective. Safety includes safety to the animal and of food products made from the treated animal. CVM also ensures that the drug’s strength, quality and purity are consistent from batch to batch and labeling is complete and truthful. The NADA process also considers impact to the environment and the safety of those who administer the drug to animals (FDA, 2015a).

The use of fenbendazole for food animals is approved under six FDA new animal drug applications (Table 3). It is dispensed over the counter. The use of ivermectin for food animals is approved under nineteen FDA new animal drug applications. It is dispensed both by veterinary prescription and over the counter (Table 3). The use of moxidectin is approved under three new drug approval applications. It is available over the counter (Table 3). The approved FDA NADA numbers for the eight additional anthelmintics approved by the FDA are provided in Table 3.

Once a NADA is approved, the FDA, under the Animal Medicinal Drug Use Clarification Act of 1994 (AMDUCA), can permit the use of the approved drug under specific conditions outside the designated or intended label use, e.g. use in species not listed in the labeling, use for indications (disease or other conditions) not listed in the labeling, use at dosage levels, frequencies, or routes of administration other than those stated in the labeling, and deviation from the labeled withdrawal time based on these different uses (FDA, 1994). This “off-label use” is only permitted in the context of a valid veterinarian-client-patient relationship and is limited to treatments when the health of an animal is threatened or suffering or death may result from failure to treat. A valid veterinarian-client-patient relationship is one in which: (1) A veterinarian has assumed the responsibility for making medical judgments regarding the health of (an) animal(s) and the need for medical treatment, and the client (the owner of the animal or animals or other caretaker) has agreed to follow the instructions of the veterinarian; (2) There is sufficient knowledge of the animal(s) by the veterinarian to initiate at least a general or preliminary diagnosis of the medical condition of the animal(s); and (3) The practicing veterinarian is readily available for follow up in case of adverse reactions or failure of the regimen of therapy. Such a relationship can exist only when the veterinarian has recently seen and is personally acquainted with the keeping and care of the animal(s) by virtue of examination of the animal(s), and/or by medically appropriate and timely visits to the premises where the animal(s) are kept (FDA, 2015b).
For example, there is not a FDA approved use for fenbendazole in domestic sheep; however, it is used under veterinary supervision for this purpose (de la Concha-Bermejillo et al., 1998). Furthermore, the National List permits the use of fenbendazole only under veterinary supervision (§ 205.603(18)(a)(i)).

There are some limitations for the AMDUCA including extralabel use of an approved new animal or human drug by a lay person (except when supervised by a veterinarian), extralabel use of an approved new animal or human drug in animal feed, extralabel use resulting in any residue that may present a risk to public health and extralabel use resulting in any residue above an established safe level, safe concentration or safe tolerance. Extralabel use of an approved new animal or human drug in food producing animals is further restricted to times when no approved animal drug with the same active ingredient is available for use or a veterinarian has found the approved animal drug ineffective, only after a diagnosis and evaluation of the conditions of the animal, after establishment of an extended withdrawal time, after assuring the maintenance of the animal’s identity and after taking appropriate measures to assure assigned time frames for withdrawal are met and no illegal drug residues occur in any food producing extralabel treated animal (FDA, 2015b).

Properties of the Substance:

Descriptions of the physical and chemical properties of all US Food and Drug Administration Center for Veterinary Medicine approved veterinary parasiticides are provided in Table 2.

Specific Uses of the Substance:

The US Food and Drug Administration Center for Veterinary Medicine and the US Department of Agriculture National Organic Program permit oral administration of fenbendazole in dairy cattle for the removal and control of lungworm (Dictyocaulus viviparous); brown stomach worm (Ostertagia ostertagi), barberpole worm (Haemonchus contortus and H. placei), small stomach worm (Trichostrongylus axei), hookworm (Bunostomum phlebotomum), threaded intestinal worm (Nematodirus helvetianus), small intestinal worm (Coopera punctata and C. oncophora), bankrupt worm (Trichostrongylus colubriformis) and nodular worm (Oesophagostomum radiatum); in beef cattle (beef) for the removal and control of stomach worm (Ostertagia ostertagi) and tapeworm (Moniezia benedeni); in goats for the removal and control of stomach worms (Haemonchus contortus and Teladorsagia circumcincta); in swine for the removal and control of lungworms (Metastrongylus apri and M. pudendotectus), roundworms (Ascaris suum), nodular worms (Oesophagostomum dentatum, O. quadriroupinatum), small stomach worms (Hyostrongylus rubidus), whipworms (Trichuris suis) and kidney worms (Stephanurus dentatus) and in turkeys for the removal and control of round worms (Ascaridia dissimilis) and cecal worms (Heterakis gallinarum). Fenbendazole is sold by Merck Animal Health as Panacur® and Safe-Guard®. It is available in liquid suspension, as granules, as a paste and in blocks. Products are dispensed both by veterinarian’s prescription and over the counter, but must be used in organic production only under veterinary supervision. For swine, turkeys, and wild sheep the NADA (141-144, 140-954, 136-116, 131-675) for fenbendazole is for use in medicated feed only. Other uses for these animals are extralabel. Furthermore, the use of fenbendazole in medicated feed for domestic sheep in food production is not permitted by the FDA (2015b).

### Approved Legal Uses of the Substance:

The US Food and Drug Administration (FDA) regulates veterinary drugs. A new animal drug is defined, in part, as any drug intended for use in animals other than man, including any drug intended for use in animal feed but not including the animal feed, the composition of which is such that the drug is not generally recognized as safe and effective for the use under the conditions prescribed, recommended, or suggested in the labeling of the drug (21 U.S.C. § 321(v)). As mandated by the Federal Food, Drug, and Cosmetic Act, a new animal drug may not be sold into interstate commerce unless it is the subject of an approved new animal drug application (NADA), abbreviated NADA (ANADA), or there is a conditional approval (CNADA) in effect pursuant to 21 U.S.C. § 360ccc or there is an index listing in effect pursuant to 21 USC § 360ccc-1 (21 U.S.C. §§ 331(a) and 360b(a)). FDA approved new drug application numbers (NADA) for parasiticides and an overview of information available at Animal Drugs@FDA for livestock parasiticides is provided in Table 3.

### Action of the Substance:

Effective veterinary parasiticides have selective toxic effects against nematode worms, i.e., kill the worm, allow the host to evacuate the worms and leave the host safe and healthy. This is true for fenbendazole, ivermectin and moxidectin which act selectively by binding to nematode β-tubulin in the case of fenbendazole and potentiating the glutamate-gated chloride (GLUCL) channel in the cases of ivermectin and moxidectin (Table 3). Binding β-tubulin disrupts the nematode digestive system and prevents egg formation, while potentiating the GLUCL channel causes spastic paralysis.

Fenbendazole, ivermectin and moxidectin work very well for susceptible parasites. However, some worms have a natural mechanism that causes subtle mutations in the genes for the β-tubulin and ion channel proteins targeted by these anthelmintics. This allows the worms in subsequent generations to avoid drug binding and enables drug resistance. Parasiticide resistance management has become an important issue in animal health. Increased use of anthelmintics in livestock production may lead to subsequent selection and increased parasiticide resistance (Xu et al., 1998; James et al., 2009). As a result, if resistance to one drug occurs, then other drugs with the same mode of action or binding site will also be ineffective. It is important to consider parasiticide mode of action in anthelmintic selection, to choose the most effective therapeutic drug (Martin, 1997).

The eleven drugs approved by the FDA for anthelmintic use in food producing animals and their modes of action, (1) nicotinic agonists, (2) γ-amino-butyric acid (GABA) agonists, (3) glutamate-gated chloride receptor potentiators and (4) microtubule blockers, are listed in Table 3.

### Combinations of the Substance:

Moxidectin is sold as Cydectin. Cydectin 1% for subcutaneous injection contains the excipients benzyl alcohol, polysorbate 80, propylene glycol, butylated hydroxytoluene, disodium edentate dehydrate, anhydrous sodium phosphate sodium acid phosphate monohydrate and water for injections. Cydectin

---

**Table 3:**

<table>
<thead>
<tr>
<th>Parasiticides: Fenbendazole, Ivermectin, Moxidectin</th>
<th>Livestock</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chabertia, Trichuris ovis, lungworms: Dictyocaulus filaria and all larval stages of the nasal bot Oestrus ovis.</td>
<td></td>
</tr>
<tr>
<td>Ivermectin is marketed by Merial, Inc. and other companies under a number of pharmaceutical labels. It is available as a drench, in liquid solution, for medicated feed, as a sustained release bolus and as a paste. Products are dispensed both by veterinarian’s prescription and over the counter.</td>
<td></td>
</tr>
<tr>
<td>Approved Legal Uses of the Substance:</td>
<td></td>
</tr>
<tr>
<td>The US Food and Drug administration (FDA) regulates veterinary drugs. A new animal drug is defined, in part, as any drug intended for use in animals other than man, including any drug intended for use in animal feed but not including the animal feed, the composition of which is such that the drug is not generally recognized as safe and effective for the use under the conditions prescribed, recommended, or suggested in the labeling of the drug (21 U.S.C. § 321(v)). As mandated by the Federal Food, Drug, and Cosmetic Act, a new animal drug may not be sold into interstate commerce unless it is the subject of an approved new animal drug application (NADA), abbreviated NADA (ANADA), or there is a conditional approval (CNADA) in effect pursuant to 21 U.S.C. § 360ccc or there is an index listing in effect pursuant to 21 USC § 360ccc-1 (21 U.S.C. §§ 331(a) and 360b(a)). FDA approved new drug application numbers (NADA) for parasiticides and an overview of information available at Animal Drugs@FDA for livestock parasiticides is provided in Table 3.</td>
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<td></td>
</tr>
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<td></td>
</tr>
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<td></td>
</tr>
<tr>
<td>The eleven drugs approved by the FDA for anthelmintic use in food producing animals and their modes of action, (1) nicotinic agonists, (2) γ-amino-butyric acid (GABA) agonists, (3) glutamate-gated chloride receptor potentiators and (4) microtubule blockers, are listed in Table 3.</td>
<td></td>
</tr>
<tr>
<td>Combinations of the Substance:</td>
<td></td>
</tr>
<tr>
<td>Moxidectin is sold as Cydectin. Cydectin 1% for subcutaneous injection contains the excipients benzyl alcohol, polysorbate 80, propylene glycol, butylated hydroxytoluene, disodium edentate dehydrate, anhydrous sodium phosphate sodium acid phosphate monohydrate and water for injections. Cydectin</td>
<td></td>
</tr>
</tbody>
</table>
### Table 3. FDA approval for Anthelmintics in use in the United States for Livestock*

<table>
<thead>
<tr>
<th>Active Ingredient: Species</th>
<th>Manufacturer(s)-Trade Name***</th>
<th>NADA-Numbers**</th>
<th>Mode of Action***</th>
</tr>
</thead>
<tbody>
<tr>
<td>Albendazole: Cattle, Sheep, Goats</td>
<td>Zoetis-Valbazen</td>
<td>110-048, 128-070, 140-934</td>
<td></td>
</tr>
<tr>
<td><strong>Fenbendazole:</strong> Cattle, Swine, Wild Sheep (Ovis), Turkeys, Goats, Deer</td>
<td>Intervet (Merck)-Panacur®, Safe-Guard®, Lincomix; Virbac-Purina Worm-A-Rest Litter Pack; Zoetis-BMD®/Safe-Guard®</td>
<td>128-620, 131-675, 132-872, 136-116, 137-600, 139-189, 140-954, 141-144</td>
<td>Nicotinic Agonists: Selectively bind to the synaptic and extrasynaptic nicotinic acetylcholine receptors on nematode muscle cells producing contraction and spastic paralysis.</td>
</tr>
<tr>
<td>Morantel tartrate: Cattle, Goats, Swine</td>
<td>Phibro Animal Health Corp.-Rumatel; Zoetis, Inc.-Rumatel, Paratect Flex</td>
<td>092-444, 093-903, 134-779</td>
<td></td>
</tr>
<tr>
<td>Doramectin: Cattle, Swine</td>
<td>Zoetis-Deectomax</td>
<td>141-061, 141-061</td>
<td></td>
</tr>
<tr>
<td>Epincomectin: Cattle</td>
<td>Merial Ltd.-Eprinex, Longrange</td>
<td>141-079, 141-327</td>
<td></td>
</tr>
<tr>
<td>Moxidectin: Cattle, Sheep</td>
<td>Boehringer Ingelheim Vetmedica, Inc.-Cydectin,</td>
<td>141-099, 141-220, 141-247</td>
<td></td>
</tr>
<tr>
<td>Piperazine: Chickens, Swine, Turkeys</td>
<td>Fleming Laboratories-Pig Wormer, Wazine</td>
<td>010-005</td>
<td>Y-Amino Butyric Acid Agonist: Selectively binds to the nematode γ-amino butyric acid receptors increasing the opening of muscle membrane chloride channels. Hyperpolarizes the membrane potential and produces spastic paralysis.</td>
</tr>
</tbody>
</table>

*anthelmintic drugs approved by the FDA for use in livestock, links are provided for fenbendazole, ivermectin and moxidectin products. Others can be found at Animal Drugs@FDA.**Animal Drugs@FDA (2015)***Martin, 1997.
-Pour On is formulated with Aromatic 100 solvent. Aromatic 100 solvent is composed of solvent naptha (petroleum), CAS #64742-95-2. This product potentially contains the toxic compounds, cumene (<1.1%), pseudocumene (<32%) and xylenes (< 2.2%).

Ivermectin is sold as Ivomec for injection. This product contains the excipients glycerol formal and propylene glycol. The pour on ivermectin product contains the excipients trolamine, crodamol CAP and isopropyl alcohol.

Fenbendazole is sold a Panacur and Safe Guard. The orally administered product contains polysorbate 80, simethicone emulsion 30%, benzyl alcohol and purified water. Febendazole paste contains the excipients carbone homopolymer type B (Allyl pentaerythritol crosslinked), propylene glycol, glycerin, sorbitol, sodium hydroxide, water, methylparaben and propylparaben.

All of the FDA livestock approved parasiticides are synthetically produced substances shown by experimental and clinical studies to be safe for application to food animals. The excipients are usually United States Pharmacopoeia (USP) grade chemicals and also subject to FDA approval.

The use of parasiticides in organic production is strictly confined to emergencies and the practice of returning livestock production to a healthy steady state that does not include the routine use of parasiticides. The current allowance of three parasiticides covering only two modes of action does not address issues of uncontrolled infection when a parasiticide fails to be effective. Combinations of parasiticides and the availability of anthelmintics with all four modes of action are considered in conventional livestock production when addressing infection and the development of anthelmintic resistance (Sargison, 2014; Bath, 2014; Taylor, 2013; Dolinska et al., 2013; McArthur and Reinemeyer, 2014; Leathwick, 2013; Busin et al., 2013; Leathwick, 2014; Le Jambre et al., 2010; Epe and Kaminsky, 2013; Leathwick and Besier, 2014; Bartram et al., 2012; Bartram, 2013).

**Status**

**Historic Use:**

Parasitism may be the weakest link in organic livestock production (Karreman, 2004). Outbreaks of disease due to nematode parasites can happen even in well managed flocks. When changes in a production system occur as a result of land use, weather, or transient exposure of susceptible animals to parasites the natural imbalance favors parasite infestation. When unnoticed, undetected and without treatment parasite infestation can lead to disease and potentially death (Stockdale, 2008). The objective of a pest control program in organic farming is to use deworming treatments only in emergencies regardless of whether the treatment is administered with natural products or not (Duval, 1997). This has not been the case with conventional farming where continuous use of parasiticides has resulted in manifold anthelmintic resistance. Anthelmintics were originally described as medicines that “kill or expel parasites from their various locations in the body.” They were divided into the vermilicides (did not kill the worms) and the vermicides (killed the worms). The areca-nut from the palm and male fern root, both natural treatments were among the first effective anthelmintics (Hoare, 1896).

During the 1920s, interest in veterinary pharmaceutical drugs, particularly anthelmintics increased prompting the discovery and development for marketing of Antimosan, Ascaridole and Avomin by Bayer. Antimosan was to be used for lungworms in cattle, Ascaridole for ascarids of pigs and Avomin for chickens. Bayer introduced levamisole in 1966, pyrantel in 1983 and ivermectin in 1997 (Harder, 2002).

Food security is the sustainable production of sufficient amounts of high quality, affordable and safe food required to underpin health and well-being of human populations worldwide (Fitzpatrick, 2013). Many aspects of livestock production including organic production have already moved from rural to peri-urban and urban settings. This change and the growing expectation for “sustainable intensification,”i.e. producing more food from less land, accompanied by more diligent land use are confounding principles for organic livestock production when parasites are considered. Much information is now known about the nematodes, their anatomy, morphology, life cycles, pathogenesis and epidemiology. Not as much is known about their ecology, but this body of research is also growing. Increasing parasiticide resistance spurred a strong movement toward understanding its underpinning molecules and mechanisms improving...
diagnostics, epidemiology and management of flocks and herds, while research into alternative approaches
to disease control, including genetic selection for resistant or resilient hosts, and vaccination, continues
(Fitzpatrick, 2013).

**Organic Foods Production Act, USDA Final Rule:**
The three parasiticides currently allowed for use by the National Organic Program in organic livestock
production as medical treatments are (i) fenbendazole (CAS # 43210–67–9)—only for use by or on the lawful
written order of a licensed veterinarian, (ii) ivermectin (CAS # 70288–86–7) and (iii) moxidectin (CAS # 113507–
06–5)—for control of internal parasites only (§ 205.603). Their use is prohibited in slaughter stock, but allowed for
emergency treatment of dairy and breeder stock when the producer’s approved preventive management system
does not prevent infestation. Milk or milk products from a treated animal cannot be labeled as organically
produced for 90 days following treatment. In breeder stock, treatment cannot occur during the last third of
gestation if the progeny will be sold as organic and cannot be used during the lactation period for breeding stock.

**International**

**Canada** -
The Canadian Organic Production Systems General Principles and Management Standards (CAN/CGSB-
32.310-2006) generally prohibit the use of parasiticides with exceptions. If no alternative treatment exists a
parasiticide may be administered under veterinary supervision as directed by the standard and mandated
by law. Treated livestock with a withdrawal period equivalent to double the label requirement or 14 days,
whichever is longer is still considered organic. Organic status for chronically infected animals is
 discontinued. The Canadian Organic Standard requires organic livestock operations to have a
comprehensive plan to minimize parasite problems in livestock, including monitoring and emergency
measures. Normally, parasiticides cannot be administered to meat, dairy or laying animals, but in
emergencies, production operations can use them: (1) if parasites are detected, (2) under veterinary
instructions, (3) with double the label withdrawal time or 14 days whichever is longer, (4) with one
treatment for slaughter animals under one year and two treatments for older animals (requiring more
treatments will lose organic status), (5) but dairy animals requiring more than two treatments lose organic
status and require a 12 month transition, (6) but dairy animals cannot be organic for slaughter, (7) and a
dam may be treated during gestation, (8) and poultry flocks can be treated, but laying hens with more than
one treatment per 12 months lose organic status and (9) the operator must provide a written action plan
with amendments to the parasite control plan.

**CODEX Alimentarius Commission, Guidelines for the Production, Processing, Labelling and Marketing
Codex Alimentarius guidelines GL 32-1999, Guidelines for the production, processing, labelling and
marketing of organically produced foods permits the use of parasiticides where specific disease or health
problems occur, or may occur, and no alternative permitted treatment or management practice exists.
Phytotherapeutic, homeopathic or ayurvedic products and trace elements are preferred to chemical
allopathic veterinary drugs or antibiotics, provided that their therapeutic effect is effective for the species of
animal and the condition for which the treatment is intended. If these are not effective in combating illness
or injury, parasiticides may be used under the responsibility of a veterinarian. Withdrawal periods should
be the double of that required by legislation with, in any case, a minimum of 48 hours. The use of
parasiticides for preventative treatments is prohibited.

Preventive use of chemically-synthesized allopathic medicinal products is not permitted in organic
farming. However, in the case of a sick animal requiring an immediate treatment, the use of chemically
synthesized allopathic medicinal products is limited to a strict minimum. Doubling withdrawal periods
after use of chemically synthesized allopathic medicinal products is suggested to guarantee the integrity of
organic production for consumers. Because widespread animal diseases would seriously affect organic
production, measures may be taken to ensure maintenance of farming or reestablishment of farming with
nonorganic animals or non-organic for a limited period in the affected areas.

**Japan Agricultural Standard (JAS) for Organic Production** –
The Japan Agricultural Standard (JAS) for Organic Production emphasizes that disease shall be prevented by strengthening resistance to disease and prevention of infestation through livestock dependent husbandry practices without unnecessary suffering. In cases where disease occurs or may occur and there is no alternative permitted treatment or management practice or laws and ordinances provide, veterinary drugs can be used. Parasiticides may only be used on livestock for the therapy purpose. In cases where parasiticides are licensed according to the Ministry Ordinance of Regulation on Use of Veterinary Drugs (Ministry Ordinance No. 42, Ministry of Agriculture, Forestry, and Fisheries (MAFF), 1980), the withdrawal period is twice the specified time. In cases where parasiticides are not licensed by MAFF, the withdrawal period is 48 hours prior to slaughter for foods, milking, and egg collection or twice the period of drug withdrawal (the period from the last administration of drugs to slaughter for foods, milking, or egg collection) defined for approval of drugs, change of approvals, reexamination of drugs, and drug efficacy review by Article 14-1, 9, 4, and 6 of the Pharmaceutical Law of Japan, whichever the longer. No specific anthelmintics are specified.

International Federation of Organic Agriculture Movements (IFOAM) –
http://www.ifoam.org/standard/norms/cover.html

Use of synthetic allopathic anthelmintics will cause an animal to lose its organic status, although producers cannot withhold such medication where doing so will result in unnecessary suffering of the livestock. An exception is included, and an animal can retain its organic status if the operator can demonstrate treatment is in compliance with IFOAM preventive animal husbandry practices, and natural and alternative medicines and treatments are unlikely to be effective to cure sickness or are not available to the operator, and the chemically synthesized allopathic veterinary medical products or antimicrobials are used under the supervision of a veterinarian, withdrawal periods are not less than double the withdrawal period required by legislation, or a minimum of 14 days, whichever is longer. The exception is granted for a maximum of three courses of remedial treatments within 12 months, or one course of treatment if the productive lifecycle of the animal is less than one year. Prophylactic use of any synthetic allopathic veterinary drug is prohibited. Vaccinations are allowed only when an endemic disease is known or expected to be a problem in the region of the farm and where this disease cannot be controlled by other management techniques, or when a vaccination is legally required.

IFOAM requires documentation of the impact of the parasiticide on the communities where they are made and used, whether the use of the substance favors any economic structure and scale, and the historical use of the substance in traditional foods. IFOAM also requires that consumer perceptions of the compatibility of inputs be taken into account, that inputs should not meet resistance or opposition of consumers of organic products, that there is scientific certainty about the impact of the substance on the environment or human health, that inputs respect the general opinion of consumers about what is natural and organic, that inputs used for animal feed and livestock production are evaluated for their impact on animal health, welfare, and behavior, that medications alleviate or prevent animal suffering and that inputs causing suffering or having a negative influence on the natural behavior or physical functioning of farm kept animals are prohibited or restricted.

Evaluation Questions for Substances to be used in Organic Livestock Production

Evaluation Question #1: Indicate which category in OFPA that the substance falls under: (A) Does the substance contain an active ingredient in any of the following categories: copper and sulfur compounds, toxins derived from bacteria; pheromones, soaps, horticultural oils, fish emulsions, treated seed, vitamins and minerals; livestock parasiticides and medicines and production aids including netting, tree wraps and seals, insect traps, sticky barriers, row covers, and equipment cleansers? (B) Is the substance a synthetic inert ingredient that is not classified by the EPA as inerts of toxicological concern (i.e., EPA List 4 inerts) (7 U.S.C. § 6517(c)(1)(B)(ii))? Is the synthetic substance an inert ingredient which is not on EPA List 4, but is exempt from a requirement of a tolerance, per 40 CFR part 180?

The livestock anthelmintics, fenbendazole, ivermectin and moxidectin fall under the Organic Foods Production Act category “livestock parasiticides” (7 U.S.C. § 6517(c)(1)(B)(ii)). The National List provides
for the use of livestock parasiticides in an organic farming operation. Three parasiticides are included in the National List: ivermectin, moxidectin and fendbendazole (7 CFR § 205.603(a)(18)).

Table 4. Methods of Synthesis for Anthelmintics approved for use in the United States for Livestock*

<table>
<thead>
<tr>
<th>Chemical Group</th>
<th>Active Ingredient</th>
<th>Manufacturer(s) **</th>
<th>Methods of Synthesis*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Benzimidazoles</td>
<td>Thiabendazole</td>
<td>Merial Ltd., ADM Alliance Nutrition, Inc.</td>
<td>Benzimidazoles are prepared chemically using a condensation of o-phenylenediamine or o-nitroaniline with a carboxylic acid derivative. N-arylamide hydrochlorides can also be transformed to benzimidazoles with sodium hypochlorite and base. (Brown et al., 1961; Grenda et al., 1965; Loewe et al, 1976).</td>
</tr>
<tr>
<td>Albendazole</td>
<td>Zoetis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fenbendazole</td>
<td>Intervet (Merck), Virbac, Zoetis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Imidazothiazoles</td>
<td>Levamisole</td>
<td>Zoetis, Intervet (Merck), Agri Laboratories, Cross Vetpharm Group, Ltd.</td>
<td>Levamisole is chemically synthesized through a number of steps. The racemic form was prepared using phenacyl bromide (Raeymakers et al., 1966). More recently, a highly enantioselective synthesis of levamisole has been accomplished by employing (R)-3-acetoxy-3-phenylpropanenitrile and (R)-3-hydroxy-3-phenylpropanenitrile obtained by both enzymatic transesterification and hydrolysis processes (Kamal et al., 2005).</td>
</tr>
<tr>
<td>Tetrahydropyrimidines</td>
<td>Morantel tartrate</td>
<td>Phibro Animal Health Corp., Zoetis, Inc.</td>
<td>The chemical name of morantel tartrate is 1,4,5,6-tetrahydro-l-methyl-2-[trans-2-(3-methyl-2-thienyl)vinyl]pyrimidine hydrogen tartrate. Synthesis of morantel involves the condensation of 3-methylthiophene-2-carbaldehyde with 1,4,5,6-tetrahydro-1,2-dimethylpyrimidinien the presence of methyl formate (Addison et al., 1974).</td>
</tr>
<tr>
<td>Pyrantel</td>
<td>Phibro, Inc., Virbac AH, Inc., ADM Alliance Nutrition, Inc., Virbac AH, Inc.</td>
<td>1,4,5,6-tetrahydro-1-methyl-[trans-(2-thienyl)vinyl]-pyrimidine, a derivative of tetrahydropyrimidine is made from 3-(2-thienyl)-acrylonitrile. 3-(2-thienyl)-acrylonitrile in a Knoevangel condensation of furfural with cyanoacetic acid. Acid hydrolysis of this compound makes 3-(2-thienyl)acrylamide. Reacting this product with propansulfone gives an iminoester which when reacted with N-methyltrimethyleyldiamine gives pyrantel (Vardanyan and Hruby, 2006).</td>
<td></td>
</tr>
<tr>
<td>Macroyclic lactones</td>
<td>Ivermectin</td>
<td>Merial Ltd., Bayer HealthCare LLC, Animal Health Division, Norbrook Laboratories Ltd, Cross Vetpharm Group Ltd., First Priority, Inc., SmartVet USA, Inc., Norbrook Laboratories Ltd., Sparhawk Laboratories, Inc.</td>
<td>The first macrocyclic lactone to be discovered and isolated was Streptomycin. It was extracted directly from Streptomyces spp. culture medium (Addinal, 1945). Ivermectin is a semi-synthetic chemically reduced 22,23-dihydro derivative of abamectin (Campbell et al., 1983). Doramectin was initially isolated through a process called “mutational biosynthesis” (Goudie et al., 1993; Dutton et al., 1990). Eprinomectin was developed by screening a large number of synthetic ivermectin analogs (Shoop et al., 1996). Milbemycins were first identified as macrocyclic lactones and isolated from cultures of Streptomyces lugrosopicosus. Thirteen were initially purified and characterized (Takiguchi et al., 1980). Moxidectin is a derivative of nemadectin. Nemadectin, a milbemycin is a Streptomyces cyanogriseus fermentation product (Asato and France, 1990). Moxidectin is related to ivermectin, but lacks a disaccharide moiety and has an O-methyl substituent at the 23-position (Deng et al., 1991).</td>
</tr>
<tr>
<td>Doramectin</td>
<td>Zoetis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eprinomectin</td>
<td>Merial Ltd.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Moxidectin</td>
<td>Boehringer Ingelheim Vetmedica, Inc.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Piperazines</td>
<td>Piperazine</td>
<td>Fleming Laboratories</td>
<td>Piperazine is synthesized from ethanolamine by heating it in ammonia at 150-220°C and 150-250 atmospheres of pressure (Vardanyan and Hruby, 2006).</td>
</tr>
</tbody>
</table>

*anthelmintic drugs approved by the FDA for use in livestock
**FDA, 2012
***Animal Drugs@FDA (2015)
Fenbendazole is an anthelmintically active 2-carboxyl-amino-benzimidazole-5(6)-phenyl ether (benzimidazole carbamate). Production of fenbendazole is described in US Patent 3954791 (Loewe et al., 1976; Table 4).

The precursor for ivermectin, avermectin B₁ is naturally produced by Streptomyces avermilitis strain that was mutagenized with high energy ultraviolet light. Hydrogenation of avermectin B₁ for 20 hours with Wilkinson's catalyst in benzene or toluene at 25°C under 1 atmosphere of hydrogen produces 85 percent 22,23-dihydroavermectin B₁ together and 3 percent of 3,4,22,23-tetrahydroavermectin B₁, 22, 23-dihydroavermectin B₁, containing at least 80 percent of 22,23-dihydroavermectin B₁a and not more than 20 percent of 22,23-dihydroavermectin B₁b, is assigned the name ivermectin (Campbell et al., 1983). The UV mutagenized Streptomyces sp., renamed Streptomyces cyanogriseus is described in a patent for the production of ivermectin that was filed in 1990 (Asato and France, 1990; Table 4).


Table 4 provides an overview of the synthetic processes involved in producing all eleven parasiticides approved by US Food and Drug Administration (FDA) for use in livestock for food production. Because all veterinary drugs must be approved by the FDA, their manufacture is an aspect of production overseen by the US federal government. The FDA provides guidance for inspection of sterile drug manufacturers (FDA, 2014a; 2014b). The FDA Center for Veterinary Medicine has published a number of guidelines focused on the new drug approval process. Some of these publications focus on anthelmintic drugs, and the manufacturing, processing or holding active of pharmaceutical ingredients.

Veterinary diagnostic tests are in development to determine whether parasites are anthelmintic resistant (Pena-Espinoza, 2014). These tests for infested livestock when available to producers will be regulated by the US Department of Agriculture, Animal Plant Health Inspection Service, National Veterinary Services Laboratory. The National Animal Health Monitoring System (NAHMS) is currently collecting data to estimate the prevalence of gastrointestinal parasites and anthelmintic resistance in sheep and cattle. Showing that one drug should be used in treatment over another in an emergency situation will provide an important tool in parasite management (Gilleard and Beech, 2007; Beech et al., 2011; Tyden et al., 2014).

**Evaluation Question #3:** Discuss whether the petitioned substance is formulated or manufactured by a chemical process, or created by naturally occurring biological processes (7 U.S.C. § 6502 (21)).

Parasiticides approved for use by the US Food and Drug administration (FDA) are manufactured synthetically with starting materials originating from the petroleum, mining or agriculture sector or as chemically modified products of bacterial (mostly Streptomyces spp.) fermentation.

**Evaluation Question #4:** Describe the persistence or concentration of the petitioned substance and/or its by-products in the environment (7 U.S.C. § 6518 (m) (2)).

Fenbendazole is insoluble in water and excreted after administration in feces. Because it is not soluble, there is little mobility of fenbendazole in soils, and a low risk of groundwater contamination. Laboratory tests show that radiolabeled fenbendazole is degraded with a half-life of 54 days. Although photo-degradation plays a role, degradation of fenbendazole in soil appears to be microbiologically dependent rather than photodegradative (Kreuzig et al., 2007).

Ivermectin is rapidly adsorbed to soil and sediment. Up to 98% of the administered dose of ivermectin may be excreted as non-metabolized drug in feces (Horvat et al., 2012). Ivermectin does not appreciably leach from soil sediment (Krogh et al., 2008). Radio-chromatographic studies have shown the ivermectin half-life for degradation to be 127 days in soil and less than 6 hours in water (Prasse et al., 2009). The environmental burden on fields manured with feces from ivermectin treated animals ranges from 0.001 to 0.09 parts per billion (ppb) depending on animal species (Halley et al., 1989).

Excretion of moxidectin is primarily through the manure of treated cattle. It is very lipophilic and not very soluble in water. Moxidectin in feces peaks at 349 ppb, 2 days after treatment and decreases to less than 10 ppb by 37 days after treatment. Feces from cattle contain no detectable levels of moxidectin thirty seven days after treatment. The half-life for degradation of moxidectin in the environment may be up to 130 days.
The environmental burden on fields manured with feces from moxidectin treated animals ranges is estimated at 0.526 parts per billion (ppb) for cattle (Fort Dodge Animal Health, 2001).

### Table 5 Environmental Persistence and Concentration of Parasiticides

<table>
<thead>
<tr>
<th>Drug</th>
<th>Environmental Impact*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thia bendazole, Albendazole, Fenbendazole</td>
<td>Thia bendazole’s affinity for binding to soil particles increases with increasing soil acidity. It is highly persistent. The field half-life for thia bendazole has been reported as 403 days. Due to its binding and slight solubility in water, it is not expected to leach readily from soil. The benzimidazoles are generally insoluble in water and sticks to humic material in terrestrial and aquatic environments. They are readily photodegradable. Benzimidazoles are introduced into the environment when they are excreted by treated animals. It is expected that 100% of the administered dose is excreted within 7 days. On a conventional 10 animal per acre cattle farm, with an expected dosage of approximately 3.5 grams per animal per treatment, and three treatments per year, the amount of benzimidazole excreted onto one acre is about 110 per year. Because the benzimidazoles stick to humic material they are not expected to run off into aquatic environments, and because they are photodegradable benzimidazoles are not expected to persist in the environment.</td>
</tr>
<tr>
<td>Morantel tartrate</td>
<td>Morantel could not be detected (&lt; 0.05 microgram/ml) in the plasma of cattle or goats following the oral administration of morantel tartrate at a dose rate of 10 mg/kg bodyweight. Morantel is difficult to detect in the milk of lactating goats, but has been detected at a concentration of 0.092 microgram/ml at 8 h after drug administration. Morantel could be detected at a concentration of 96 +/- 4.5 micrograms/g (dry weight) in the feces of a calf 24 h after treatment with 10 mg/kg bodyweight of morantel tartrate. The concentration of morantel in replicate samples of feces exposed to natural atmosphere, but not to soil or soil organisms, declined slowly over the following 322 days. At day 322 after the start of the experiment 8.8 micrograms/g of morantel could be measured in the remaining fecal material. Throughout a fecal degradation study the concentration of morantel in the crusts of replicate sample pats was lower than the concentration in the core samples. This is the result of photodegradation. Morantel is not active against bacteria or fungi. It is degraded in the soil. Pyrantel and morantel are chemically related. Persistence of either is not expected in the environment.</td>
</tr>
<tr>
<td>Pyrantel</td>
<td>Levamisole is highly soluble in water. Thus it can leak into the aquatic environment via runoff. Levamisole may non-enzymatically decompose to form three degradation products. The decomposition is temperature and pH dependent. Storage for a period of time under relatively mild, neutral and alkaline conditions causes degradation into three products one of which is responsible could be responsible for the immunomodulatory activity.</td>
</tr>
<tr>
<td>Ivermectin</td>
<td>Avermectins are excreted mainly through feces as non-metabolized drug, and their excretion profile depends strongly on the drug formulation, dosage, animal species, and sex of the animal. The fecal excretion of doramectin was studied for 56 days in treated female and castrated cattle and found that the excretion was approximately 38%, with the maximum excretion levels appearing 21 days after treatment. A similar time profile was observed for abamectin and doramectin excretion in sheep feces, observing maximum levels in the first days after treatment. Pigs excrete the highest levels of doramectin in the feces in the early days after treatment, although doramectin could still be detected in the feces after 60 days. In the field experiment, the application of manure containing doramectin under the specified conditions led to the presence of low levels (&lt;5 ng/g) of the drug in the soil. Seven months after the manure application, traces of doramectin were still detected from the surface of the soil to a 90 cm depth. Successive applications of manure from pigs treated with doramectin in a specific area would produce an accumulation of this drug in the soil that reached toxic levels for soil fauna. Ivermectin and moxidectin have been evaluated for their toxicity to insects, particularly those involved in compost production. Both were found to be toxic to these animals. Eprinomectin is used for treatment of parasites in cattle, including lactating cows. The recommended dosage is a single dose of 0.5 mg kg⁻¹ b.w. applied topically along the midline of the animal’s back. Eprinomectin, a drug with high efficacy and a large safety margin for mammals, is mainly excreted in the bile and feces; only a small proportion is excreted in the urine or is present in milk. During the 28 days after topical application of 0.5 mg kg⁻¹ b.w. radiolabelled eprinomectin to 8–10 month old calves only 0.35% of the applied dose was found in the urine whereas 17 to 19.8% was found in the feces. Eprinomectin B1a was the most abundant residue in the feces, representing 78.3% of total residues. During the 14 days after drug administration the amount of radioactivity present in the milk represented 0.32 to 0.54% of the drug. Moxidectin is excreted in feces. It is the least toxic to dung beetles of the macrocyclic lactone anthelmintics. Moxidectin is both microbially and photo-degraded in dung pats in the soil.</td>
</tr>
<tr>
<td>Doramectin</td>
<td>In pigs, piperazine is readily absorbed from the gastrointestinal tract, and the major part of the resorbed compound is excreted as unchanged piperazine during the first 48 hours. The principal route of excretion of piperazine and its metabolites is via urine, with a minor fraction recovered from feces (16 %). Piperazine can be assumed to be rapidly photolysed in the atmosphere, the half-life was calculated to be 0.8 hours. In natural water it is considered to be stable towards photolysis. From non-standard studies it can be expected that piperazine is hydrolytically stable under environmentally relevant conditions. Piperazine is not readily biodegradable but can be considered to be inherently degradable.</td>
</tr>
</tbody>
</table>

*mg-milligrams, kg-kilograms, b.w.- body weight, g-grams, ng-nanogram, ml-milliliter, cm-centimeter

1 EPA, 2002
2 US Food and Drug Administration, 1995
3 Phoenix Scientific, 2002
4 Horvat et al., 2012
5 McKellar et al., 1993
6 Pfizer, 1979
7 Gil-Diaz et al., 2011
8 Blanckenhorn et al., 2012
9 Nenka et al., 2007
10 OECD, 2004
Table 6 provides an overview of the environmental fate of the parasiticides. Most reports on the environmental persistence of the parasiticides reflect continuous use for prevention and treatment.

**Evaluation Question #5:** Describe the toxicity and mode of action of the substance and of its breakdown products and any contaminants. Describe the persistence and areas of concentration in the environment of the substance and its breakdown products (7 U.S.C. § 6518 (m) (2)).

<table>
<thead>
<tr>
<th></th>
<th>Agricultural Soils</th>
<th>Prairie Soils</th>
<th>Forest Soils</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Bacteria</strong></td>
<td>100 million to 1 billion</td>
<td>100 million to 1 billion</td>
<td>100 million to 1 billion</td>
</tr>
<tr>
<td><strong>Fungi</strong></td>
<td>Several diverse isolates. (Dominated by vesicular arbuscular mycorhizal fungi).</td>
<td>Tens to hundreds of diverse isolate. (Dominated by vesicular arbuscular mycorhizal fungi).</td>
<td>Several hundred diverse isolates in deciduous forests. One to forty miles in coniferous forests (dominated by ectomycorrhizal fungi).</td>
</tr>
<tr>
<td><strong>Protozoa</strong></td>
<td>Several thousand flagellates and amoebae, one hundred to several hundred ciliates.</td>
<td>Several thousand flagellates and amoebae, one hundred to several hundred ciliates.</td>
<td>Several hundred thousand flagellates and amoebae, fewer flagellates.</td>
</tr>
<tr>
<td><strong>Nematodes</strong></td>
<td>Ten to twenty bacterial feeders. A few fungal-feeders. Few predatory nematodes</td>
<td>Tens to several hundreds</td>
<td>Several hundred bacterial and fungal feeders. Many predatory nematodes.</td>
</tr>
<tr>
<td><strong>Arthropods</strong></td>
<td>Up to one hundred</td>
<td>Five hundred to two thousand</td>
<td>Ten to twenty five thousand. Many more species than in agricultural soils</td>
</tr>
<tr>
<td><strong>Earthworms</strong></td>
<td>Five to thirty. More in soils with high organic matter</td>
<td>Ten to fifty. Arid or semi-arid areas have none</td>
<td>Ten to fifty in deciduous woodlands, very few in coniferous forests.</td>
</tr>
</tbody>
</table>

*(Ingham, M.R., 1999)*

Maintaining healthy forage fields and healthy soils is important for livestock health (Brunetti and Karreman, 2006). Fields and pastures have unique soil ecologies with specific ratios of bacteria, fungi, and other microorganisms, and a particular level of complexity within each group of organisms (Table 6). These differences result from soil, vegetation, and climate factors, as well as land management practices. Grasslands and agricultural soils usually have bacterially-dominated food webs. Highly productive agricultural soils tend to have ratios of fungal to bacterial biomass near 1:1 or somewhat less. Organisms reflect their food source. For example, protozoa are abundant where bacteria are plentiful. Where bacteria dominate over fungi, nematodes that eat bacteria are more numerous than nematodes that eat fungi (Lavelle et al., 2006).

This balance influences the survival and persistence of pathogenic nematodes and their predators. Management practices change food webs. For example, in reduced tillage agricultural systems, the ratio of
fungi to bacteria increases over time, and earthworms and arthropods become more plentiful (Ingham, 1999).

These organisms are all essential in breaking down manure, particularly manure containing parasites.

Fenbendazole, ivermectin and moxidectin are very effective anthelmintics. Their residues are excreted in urine and feces, and may hinder the soil food webs from effectively breaking down manure and maintaining pasture health (Karremans, 2004). When undegraded, dung pats harbor nematodes parasitic in livestock, reduce available grazing area, and represent a loss of soil nitrogen in pastures. (Floate et al., 2005).

Fenbendazole toxicity was demonstrated in pigeons and doves leading the authors of the study to suggestion a toxic etiology for fenbendazole in birds of the order Columbiformes treatment (Howard et al., 2002).

The fate of fenbendazole in manure and manured soils has been studied under laboratory and field conditions. In pig manure, benzimidazoles disappear slowly. After a 102 day incubation period, 80% fenbendazole remains. The latter was accompanied by 4% of the corresponding metabolite fenbendazole-sulfoxide. Fenbendazole-sulfoxide remains in clay soil samples after 54 days (Kreuzig et al., 2007).

Excreted fenbendazole and ivermectin residues in cattle dung pats do not significantly affect adult dung beetles or adult dipteran flies. However, excreted ivermectin produces toxic effects on the larval development of the same dung-colonizing families of insects, while fenbendazole lacks such toxic effects (Strong et al., 1995).

Fenbendazole does not appear to hinder rapid disappearance and mineralization of cattle dung pats in pastures and does not appear to affect the role that earthworms play in this process. Excreted ivermectin does delay the disappearance of dung pats, but does not affect earthworm populations or health. The delay in ivermectin treated soils may be the result of its toxicity to insects (Svendsen et al., 2003). Ivermectin has low level toxicity to fish and aquatic life (Halley et al., 1993).

Much work has been done to study the macrocyclic lactones particularly ivermectin, and others, highlighting the effects of these parasiticides (Forster et al., 2011). Among the macrocyclic lactones, ivermectin is generally more toxic to insects than moxidectin. Little information is available regarding the effects of parasiticide residues on other soil food web microorganisms that facilitate the process of dung degradation (e.g., fungi, free-living nematodes, collembolans, mites). Residues of ivermectin and fenbendazole are toxic to the soil nematode Pristionchus maupasi at concentrations greater than 3 ppm and 10 to 20 ppm wet weight of dung, respectively, but sub-lethal concentrations may enhance the growth of the nematode in dung of treated cattle (Floate et al., 2005). Moxidectin has been shown to adversely affect the larvae of Musca autumnalis, Onthophagus gazella, Onitis alexis and Haematobia irritans, adult and larvae of Onthophagus binodis and to reduce the brood mass production of O. binodis and O. alexis (McKellar, 1997).

Harmonization of veterinary medicine testing requirements is coordinated by the International Cooperation on Harmonization of Technical Requirements for Registration of Veterinary Products (VICH). Members are the European Union (EU), Japan, and the USA, with Australia/New Zealand and Canada as observers. The VICH Ecotoxicity/Environmental Impact Assessment Working Group is developing ring-tested toxicity test methods for dung beetles and dung flies. The Dung Organism Toxicity Testing Standardization (DOTTS) Group in cooperation with VICH has developed several tests for dung fly and beetle ecotoxicity. In conjunction with VICH and the DOTTS Group, the FDA has also provided guidance for industry on assessing ecotoxicity (FDA, 2006).

The parasiticides belong to widely different chemical groups making it difficult to generalize their environmental risk. Exposures, biocidal properties and the effects of combinations of products have been or still need to be assessed for each group or individual drug. Data including persistence and adsorption in soil and manure, the influence of temperature and soil properties and specific toxicity which can range over several orders of magnitude is still being gathered for the parasiticides (Schmitt and Rombke, 2008).

Residues persisting in the dung of treated animals for days, weeks or months after treatment can adversely affect guilds of coprophilous insects, mites, nematodes, earthworms, and fungi that accelerate degradation of the dung pat. Table 7 provides an overview of toxicity resulting from the eleven anthelmintics approved by the US Food and Drug Administration.
**Evaluation Question #6:** Describe any environmental contamination that could result from the petitioned substance’s manufacture, use, misuse, or disposal (7 U.S.C. § 6518 (m) (3)).

Fenbendazole is manufactured by a process that requires petrochemicals such as benzene and various amines. These are considered toxic compounds. Fenbendazole is not generally considered toxic to humans at regulated doses (FDA, 1995).

Both ivermectin and moxidectin are produced by processes involving bacterial fermentation and subsequent chemical modification after the fermentation product is isolated. Environmental contamination as a result of the manufacture of either product is unlikely. Table 7 provides an overview of environmental persistence and toxicity for the FDA approved livestock parasiticides.

**Table 7 Environmental Toxicity of Parasiticides**

<table>
<thead>
<tr>
<th>Drug</th>
<th>Toxicity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thiaobendazole⁵</td>
<td>Thiaobendazole is toxic to species of freshwater estuarine fish and freshwater/estuarine invertebrates and practically non-toxic to birds and mammals. Birds and mammals can be exposed to pesticides applied as foliar sprays or powders by a variety of routes, including ingestion, dermal contact, and inhalation. It is not expected to appreciably accumulate in aquatic organisms, although the bio-concentration factor for thiaobendazole in whole fish is 87 times the ambient water concentrations. Fish eliminated the compound within 3 days after being placed in thiaobendazole-free water. Earthworms are sensitive to thiaobendazole (LD₅₀ = approx. 20 ug/worm), while bees are not. Administration of Allobendazole during gestation has been shown to cause embryotoxic effects in cattle, rat, rabbit and sheep. Observed effects include increase of resorptions, decreased fetal weight and increase of teratogenic effects, such as vascular, craniofacial, skeletal and external malformations. The dung from fenbendazole-treated animals has no obvious impact on the coleopteran or dipteran species encountered in this study, and the dung pats from the fenbendazole-treated animals were not consistently different from the pats of untreated animals. Earthworms are not significantly affected by fenbendazole.</td>
</tr>
<tr>
<td>Morantel tartrate⁷</td>
<td>Morantel is non-toxic for aquatic species. It is considered a substrate for microbial degradation in the soil. No adverse interactions with soil or aquatic environment have been observed. Both pyrantel and morantel are counter indicated for gestating animals. Pyrantel is permitted at 10 parts per million (PPM) in the kidney and 1 ppm in muscle. Morantel does not alter the rate of dung digestion.</td>
</tr>
<tr>
<td>Pyrantel⁸</td>
<td>Levamisole does not affect the fauna or the degradation of dung from inoculated animals. Breakdown products levamisole may be associated with immunomodulation effects.</td>
</tr>
<tr>
<td>Ivermectin⁹</td>
<td>The macrocyclic lactones can be ranked in decreasing order of toxicity to dung-dwelling insects as abamectin&gt;doramectin ≥ ivermectin &gt; eprinomectin&gt;&gt;moxidectin. Ivermectin has been shown to exhibit toxicity for certain dung-colonizing insects. Patterns of interaction are complex since some of these drugs are insect attractants as well as insecticide and some studies have not considered all of the aspects involved in short and longer term effects since insect activity is a composite measure of residue toxicity, the number and species of composition of insect colonists and mortality factors associated with the co-occurrence of species in dung. Flies that are sensitive to ivermectin are also sensitive to moxidectin.</td>
</tr>
<tr>
<td>Doramectin₁₀</td>
<td>Piperazine is rapidly photolysed in the atmosphere with a half-life of 0.8 hours. In natural water it is considered to be stable towards photolysis. Piperazine is hydrolytically stable under environmentally relevant conditions and not readily biodegradable but can be considered to be inherently degradable. There is no considerable potential for bioaccumulation; a bioconcentration factor of &lt; 3.9 for <em>Cyprinus carpio</em> is reported. Short-term effect studies on aquatic organisms are available for algae, aquatic invertebrates and fish. For algae (<em>Selenastrum capricornutum</em>) the no observed effect concentration (22 h growth inhibition test) is &gt; 1000 mg/l. For <em>Daphnia magna</em> the 48 hour 50% effective concentration for is 21 mg/l and for fish (<em>Poecilia reticulata</em>) the 96 hour 50% lethal concentration is &gt; 1800 mg/l. A long-term study for <em>Daphnia magna</em>, which is the most sensitive of the species tested in short term studies, results in no observable effect concentration (21 d semi-static reproduction study) of 12.5 mg/l.</td>
</tr>
<tr>
<td>Eprinomectin₁¹</td>
<td></td>
</tr>
<tr>
<td>Moxidectin₁¹</td>
<td></td>
</tr>
<tr>
<td>Piperazine</td>
<td></td>
</tr>
</tbody>
</table>


**Evaluation Question #7:** Describe any known chemical interactions between the petitioned substance and other substances used in organic crop or livestock production or handling. Describe any environmental or human health effects from these chemical interactions (7 U.S.C. § 6518 (m) (1)).
Fenbendazole is insoluble in water, is not a leachate, binds tightly to soil and is not expected to migrate in soil. The only route for fenbendazole to enter the environment is through animal excretion or spillage.

Fenbendazole degrades in soil through microbial and photodegradative processes, taking up to 60 days (Hoechst-Roussel Agrivet, 1995).

Ivermectin has very little solubility in water. The only route for entry into the environment is through animal excretion. Ivermectin has limited mobility in soil because it is lipophilic and tightly binds soil particles. The half-life for degradation of ivermectin in soil can be as long 240 d in natural soil depending on the soil type. Degradation in water is much faster with a half-life as short as 2.9 days. Ivermectin is hydrolytically unstable at pH 6.3. Predicted environmental concentrations based on the introduction of manure to field is relatively low and on the order of 100 parts per billion (ppb). It is toxic to fish at concentrations between 3 and 17 ppb. Generally, since its introduction no risks from appropriate use of ivermectin have been established for the environment or for human health. However, it has been consistently shown that ivermectin is unacceptably toxic for larval forms of arthropod insects (dung organisms) and daphnids (Liebig et al., 2010; Oh et al., 2006).

**Evaluation Question #8:** Describe any effects of the petitioned substance on biological or chemical interactions in the agro-ecosystem, including physiological effects on soil organisms (including the salt index and solubility of the soil), crops, and livestock (7 U.S.C. § 6518 (m) (5)).

Soil invertebrates are enormously diverse representing as much as 23% of the total diversity of living organisms. Their sizes range across three orders of magnitude: the smallest nematodes and protozoa (protists) of the microfauna less than 200 μm on average live in the water-filled porosity (Lavelle et al., 2006). The biological effect of fenbendazole, ivermectin and moxidectin on the agro-ecosystem is twofold: 1) sub-lethal or lethal toxicity for soil food guild organisms, such as dung beetles and beneficial nematodes, involved in degrading manure, processing humus and maintaining soil and forage field health and 2) selection and transmission of populations of anthelmintic resistant organisms such as nematodes to the soil that will subsequently be untreatable with fenbendazole, ivermectin or moxidectin upon reinfection. More than ninety percent of nematodes can be found in a non-parasitic or free living stage (Fiel et al., 2012).

The algorithm for treatment on organic farms includes fecal soiling/diarrhea, anemia, low weight gains and high fecal egg counts (Cabaet et al., 2009). Ivermectin and moxidectin are excreted into the environment in feces, while fenbendazole is excreted in urine and feces. In addition, the wash off of topically applied anthelmintics, spillage and inappropriate disposal provide additional routes of entry into the environment (Beynon, 2012). Healthy adult animals develop immunological tolerance to helminth parasites (Tizard, 2013). Treatment with parasiticides is necessary only for sick and very young animals (Karreman, 2004). Under treatment, resistant parasites, their eggs and residual anthelmintic drugs are shed in feces and urine returning to the soil. Coprophilous arthropods and microorganisms normally involved in dung pat disappearance avoid the treated dung pat or are killed as a result of anthelmintic treatment prolonging the survival of residual pathogens and promoting their return to soil and forage, where they are untreatable (Strong et al., 1996; Svendsen et al., 2003; McKellar, 1997). Some species of nematodes are both plant and animal pathogens (Jasmer et al., 2003). Further loss as a result of introduction of fenbendazole, ivermectin and moxidectin to dung pats and the soil, of otherwise predatory or competitive nematodes removes selective pressure against the parasites decreasing sustainability (Lavelle et al., 2004).

There are several nematode food guilds, including bacteria eaters, fungus eaters, and predatory nematodes. Any of these have the potential for parasitism. Parasiticides are not specific and beneficial nematodes may be killed by secondary excretion. Disturbing the ecosystem and eliminating respective food sources leaves the most aggressive parasite species without competitors. This is currently the most important interaction of anthelmintic drugs with the agro-ecosystem and there is still much to discover regarding interactions in soils and the multiple roles that invertebrates may play in controlling pests and diseases (Lavelle et al., 2006).

Anthelmintic drug resistance stems from the inability of the anthelmintic drug to affect specific nematode functions or anatomical changes, i.e., mode of action. Only four modes of action have been identified for anthelmintic drugs: 1) neuromuscular inhibition, 2a) ion channel inhibition: GABA-gated, 2b) GLUCL-gated and 3) β-tubulin binding/inhibition of microtubule formation. If resistance to a particular

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Parasiticides: Fenbendazole, Ivermectin, Moxidectin

Livestock

anthelmintic has occurred, it is likely that another anthelmintic with the same mode of action will also be
ineffective although other anthelmintics with another mode of action may still be effective. Table 8
provides the dates of introduction of some anthelmintic drugs and the subsequent report dates of
anthelmintic resistance.

Piperazine and morantel are GABA (γ-amino-butyric acid) agonists of receptors on nematode muscles and
causes flaccid paralysis. The macrocyclic lactones increase the opening of glutamate-gated chloride
channels and produce paralysis of pharyngeal pumping. Moxidectin was initially identified as a GABA
antagonist, but its primary anthelmintic activity was subsequently shown to be a GLUCL potentiator. The
benzimidazole drugs bind selectively to β-tubulin of nematodes and inhibit microtubule formation
(Martin, 1997).

The imidazothiazole, levamisole and the tetrahydropyrimidines, pyrantel and morantel are anthelmintics
that target the nicotinic acetylcholine gated cation-channels. These mediate fast synaptic signaling in the
nervous-musculature of nematodes acting as agonists to increase the flow of cations leading to a rigid
paralysis. These gated channels share a pentameric quaternary subunit structure in which a single subunit
can produce a homomeric channel, but more commonly different subunits combine to form a heteromeric
channel. Thus, resistance can occur as a result of subunit polymorphism, at the protein level or allele
variation at the DNA level. Deoxyribonucleic acid (DNA) sequence changes at three sites in the beta-
tubulin gene are thought to be the major cause of fenbendazole resistance. However, changes in the gene
for the drug transporter P-glycoprotein have also been linked with fenbendazole resistance. Ivermectin,
doramectin, eprinomectin and moxidectin are allosteric modulators of nematode glutamate channels and
cause an inhibition of pharyngeal pumping, motility and egg-laying. These channels are also composed of
protein subunits and may be homo- or heteromeric. Resistance to fenbendazole affects resistance to
ivermectin and moxidectin. However, the specific allele associated with fenbendazole resistance is different
from that associated with ivermectin and moxidectin resistance, the possibility of a mechanistic link
between resistance to fenbendazole, ivermectin and moxidectin suggests that selection for resistance with
one drug could alter the development of resistance to the second drug (Beech et al., 2011).

Table 8 The development of anthelmintic resistance

<table>
<thead>
<tr>
<th>Drug Class</th>
<th>Drug Name</th>
<th>Host</th>
<th>Year of Introduction</th>
<th>Year Resistance Reported</th>
</tr>
</thead>
<tbody>
<tr>
<td>Benzimidazoles</td>
<td>Thiabendazole</td>
<td>Sheep</td>
<td>1961</td>
<td>1964</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Horse</td>
<td>1962</td>
<td>1965</td>
</tr>
<tr>
<td>Ferbendazole</td>
<td></td>
<td>Sheep</td>
<td>1990</td>
<td>2011</td>
</tr>
<tr>
<td>Imidazothiazoles-Tetrahydropyrimidines</td>
<td>Levamisole</td>
<td>Sheep</td>
<td>1970</td>
<td>1979</td>
</tr>
<tr>
<td></td>
<td>Pyrantel</td>
<td>Horse</td>
<td>1974</td>
<td>1996</td>
</tr>
<tr>
<td>Macrocyclic Lactones</td>
<td>Ivermectin</td>
<td>Sheep</td>
<td>1981</td>
<td>1988</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Horse</td>
<td>1983</td>
<td>2002</td>
</tr>
<tr>
<td></td>
<td>Moxidectin</td>
<td>Sheep</td>
<td>1991</td>
<td>1995</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Horse</td>
<td>1995</td>
<td>2003</td>
</tr>
</tbody>
</table>

Adapted from James et al., 2009; Kaplan, 2004; George et al., 2011

P-glycoprotein is a large (170 kDa) integral membrane protein. It is able to transport a wide variety of
lipophilic substances, including many drugs. P-glycoprotein confers multidrug resistance (MDR) by active
transport of drugs, coupled to the binding and/or hydrolysis of ATP. This transport reduces the amount of
drug reaching its target and consequently reduces the effect of the drug. MDR drugs enter the cell by
passive diffusion and are actively extruded by the transport protein P-glycoprotein. P-glycoprotein can be
induced by drug treatment. P-glycoprotein is able to transport many different drugs and consequently
confers cross-resistance to many other drugs. The level of this cross-resistance varies and might be different
for different cells. P-glycoprotein-expressing cells might be more resistant to other drugs than to the drug
used to induce its expression (James et al., 2009).

An experimental model for the development of ivermectin-resistant strains of the model nematode
Caenorhabditis elegans through step-wise exposure to increasing doses of ivermectin commencing with a
non-toxic dose of 1 ng/ml also showed a multidrug resistance phenotype with cross-resistance to the
related drug moxidectin and to other anthelmintics, levamisole and pyrantel, but not albendazole. The
resistance phenotype was associated with increased expression of the multidrug resistance proteins (MRPs)
and P glycoproteins (James and Davey, 2009).

Evaluation Question #9: Discuss and summarize findings on whether the use of the petitioned
substance may be harmful to the environment (7 U.S.C. § 6517 (c) (1) (A) (i) and 7 U.S.C. § 6517 (c) (2) (A)
(i)).

Land use and chemical application respectively for livestock production and/or control of specific
pathogenic species potentially perturbs or destroys the habitat for many other beneficial organisms
(Rasmann, 2012; Zhou et al., 2012). A chemical prescription to kill an enemy (whipworm) of the farmer can
also lead to the destruction of a friend (Sykes, 1949). The impact and effects of prolonged use of
anthelmintic parasiticides on terrestrial ecology are not well understood. Table 7 provides an overview of
environmental toxicity for FDA approved anthelmintics.

Parasiticides used preventively are detectable in soils, surface water and groundwater. Estimates based on
animals dosage, land usage and degradation rate range from 0.01 parts per billion (ppb) to 500 or more ppb
(Oh et al., 2006; Liebig et al., 2010). Although fenbendazole, ivermectin and moxidectin have not been
found in agricultural products grown on fields manured with dung from treated animals, low
concentrations of levamisole have been detected experimentally in carrot and corn (Boxall et al., 2006).

Diversity and abundance of the soil invertebrate community, particularly the nematode population is not
affected by a shift from conventional to organic farming. However, there is a significant different between
either conventional or organic grazed pastures and unfertilized, ungrazed pasture. Physically, the pore size
of soil from the un-grazed, unfertilized pasture is large. This is likely to be due to the absence of livestock
treading on the soil. There is a considerable effect in both organic and conventional farming from the
presence of animals on the pasture, suggesting that land management practices such as stocking rate are
important in influencing nematode populations and that fallowing a pasture is important (Schon et al.,
2011a).

Organic livestock production avoids the development of anthelmintic drug resistances, through good
forage maintenance, exercise for livestock and practices limiting the use of holistic anthelmintic treatments.
Parasiticides may only be needed in emergencies where the organic production plan has failed (Lund and
Algers, 2003). High forage consumption and increased livestock grazing creates pasture heterogeneity and
potential imbalance between nutrition and parasitism for foraging livestock, particularly in the transition
from conventional farming (regular and prophylactic parasiticide use) to organic farming (no parasiticide
use). Overcoming these disturbances while converting forage fields from conventional to organic farming
requires careful attention to pasture conditions, water quality and the relationships between the organisms,
e.g. between plants and fungi and between invertebrates and gut organisms (Callaham et al., 2006; Smith et
al., 2009; Brunetti and Karreman, 2006; Perry, 1995).

Among the nematodes, larger, predatory and omnivorous nematodes are sensitive to the influence of
livestock on the soil environment. These nematodes are less abundant in grazed paddocks. While larger
nematodes are sensitive to livestock disturbance, they are abundant in mown and irrigated plots (Schon et
al., 2011b).

Especially in grasslands, nematodes have been found to play an important role in the transfer of energy
and matter through the soil food web because of their central and diverse trophic positions. Different
functional groups can be distinguished within the nematode community: nematodes may belong to the
primary consumer group (plant feeders), the secondary consumer group (bacterivores and fungivores), or
the tertiary consumer group (predators and omnivores). Management practices such as high stocking
<table>
<thead>
<tr>
<th>Drug</th>
<th>Maximum Residue Limit*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Animal Species</td>
<td>Cattle</td>
</tr>
<tr>
<td>Thiabendazole 1</td>
<td>Milk/100 µg/liter; Kidney, Muscle, Fat, Liver/100 µg/kilogram</td>
</tr>
<tr>
<td>Albendazole</td>
<td>Milk/100 µg/liter; Muscle, Fat, 100 µg/kilogram; Kidney, Liver/5000 µg/kilogram</td>
</tr>
<tr>
<td>Fenbendazole 1</td>
<td>Milk/100 µg/liter; Kidney, Muscle, Fat, Liver/100 µg/kilogram</td>
</tr>
<tr>
<td>Levamisole 1</td>
<td>Kidney, Muscle, Fat, Liver/100 µg/kilogram</td>
</tr>
<tr>
<td>Morantel tartrate 2</td>
<td>Milk/100 µg/liter; Muscle, Fat, Liver/100 µg/kilogram; Kidney/200 µg/kilogram</td>
</tr>
<tr>
<td>Pyrantel 3</td>
<td></td>
</tr>
<tr>
<td>Ivermectin 1</td>
<td>Milk/10 µg/liter; Fat/40 µg/kilogram; Liver/100 µg/kilogram</td>
</tr>
<tr>
<td>Doramectin 3</td>
<td>Milk/15 µg/liter; Muscle/10 µg/kilogram; Fat/150 µg/kilogram; Liver/100 µg/kilogram; Kidney/30 µg/kilogram</td>
</tr>
<tr>
<td>Eprinomectin 3</td>
<td>Milk/20 µg/liter; Muscle/100 µg/kilogram; Fat/250 µg/kilogram; Liver/2000 µg/kilogram; Kidney/300 µg/kilogram</td>
</tr>
<tr>
<td>Moxidectin 1</td>
<td>Muscle/20 µg/kilogram; Fat/500 µg/kilogram; Liver/100 µg/kilogram; Kidney/50 µg/kilogram</td>
</tr>
<tr>
<td>Piperazine 3</td>
<td>Muscle/0.1 µg/kilogram; 0.1 µg/kilogram; Kidney/0.1 µg/kilogram</td>
</tr>
</tbody>
</table>

1 Codex Alimentarius, 2014  2 Committee for Veterinary Medicinal Products: Morantel  3 Animal Drugs@FDA
density cause shifts in the functional groups and ultimately affect soil nutrient dynamics. Ecological modelling suggests that a strong, selective, human-induced pressure is acting on most taxa, indicating decreased ecosystem resilience (lower biodiversity within functional groups) as a result of increased management intensity. Many taxa are endangered as even cosmopolitan, unspecialized nematodes such as the Teratocephalidae (Secernentea) appear suppressed under intensive management. In organic farming systems, manuring provides a positive influence on microflora and bacterivorous nematodes such as Metateratocephalus and Teratocephalus (Mulder et al., 2003).

**Evaluation Question #10:** Describe and summarize any reported effects upon human health from use of the petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (i), 7 U.S.C. § 6517 (c) (2) (A) (ii)) and 7 U.S.C. § 6518 (m) (4)).

The no observable effect level (NOEL) for parasiticides is determined by drug manufacturer and approved by the US Food and Drug Administration, Codex Alimentarius or other national or international standard setting organization. Protocols are provided by these federal agencies that detail testing and evaluation of the drugs. The NOEL is usually determined in an animal model. The NOEL values for fenbendazole, ivermectin and moxidectin are respectively, 0.7 milligram/kilogram body weight per day (mg/kg bd/day), 1.5 mg/kg bd/day and 10 mg/kg bd/day. The NOEL is used to determine the Adult Daily Intake (ADI) or the maximum residue limit (MRL). Withdrawal time is the time that it takes for the concentration in milk, eggs and meat that will be consumed by people to drop from the residue level at administration to the ADI, MRL, or safe level. Drug side effects are provided on the respective drug label. Some maximum residue limits according for the US Food and Drug administration approved parasiticides are provided in Table 10.

Fenbendazole has been determined to be safe to human health when food derived from treated animals is ingested (FDA, 1995). In 2014, the US Food Safety Inspection Service found no violative positive meat samples containing moxidectin or ivermectin in the 2014 National Residue Program for Meat Poultry and Egg Products out of 237 samples tested. In 2011, the FSIS found 3 violations for moxidectin and 2 violations for ivermectin from 2019 samples including beef cows, boars, dairy cow, veal, goats, heavy calves, market hogs, mature sheep, roaster hogs and steer. Fenbendazole has not appeared recently in this survey, but will be surveyed in 2015.

**Evaluation Question #11:** Describe all natural (non-synthetic) substances or products which may be used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).

Naturally, livestock develops an immune response to nematodes and becomes resistant or tolerates them without signs of disease. Because calves do not have a mature immune system, they may not be able to mount an immune response upon infection. The same is also true for older and immunocompromised animals (Tizard, 2013). Worming with homeopathic and botanical remedies should begin strategically during the first autumn of life to accommodate the low body reserves expected with calves (Karreman, 2004).

Homeopathic wormers are available commercially that satisfy the organic rule. These are available as veterinary preparations with valid labeling systems so that their use may easily be audited (Brunetti and Karreman, 2006). Users of these remedies should be sure that the material has an appropriate potency and the source from which it was extracted is verified and correct. A list of natural wormers is provided in Table 11. Herbal remedies with anthelmintic properties were commonly adopted and used as a part of traditional animal husbandry. Some have not been evaluated with modern techniques, but may cause toxic side effects, however in most cases they represent a good alternative to the use of synthetic drugs (Duval, 1997). Crude drugs are not as efficient in their anthelmintic effects as synthetics, but are nonetheless effective and used among many cultures throughout the world (Mali, R. G. and Mehta, A.A., 2008).

The seeds from Chenopodium ambrosioides L. var. antherminticum A. Gray (Chenopodiaceae) also known as American wormseed are used to produce chenopodium oil (USP) (Kiuchi et al., 2002). Chenopodium oil is used as an anthelmintic treatment for hookworm and round worms. It is very effective against ascarids (Karreman, 2004). Chenopodium does not kill the worms but paralyzes them. They are expelled with a cathartic such as castor oil (Hatcher and Wilbert, 1915).
Sabina, USP is the tops of Juniperous Sabina, an evergreen shrub of Northern Europe, Asia and America. It contains oleum sabinae (volatile oil), fixed oil, gum, resin, gallic acid, chlorophyll, lignin and calcareous salts and salts of potassium (Karreman, 2004; Hare, 1904). Oleum Sabina is used as an anthelmintic. It is effective, but also inflammatory and poisonous (Hare, 1904).

Table 10 Botanical and Alternative De-wormers

<table>
<thead>
<tr>
<th>from Duval, 1997</th>
<th>from Karreman, 2004</th>
</tr>
</thead>
<tbody>
<tr>
<td>Garlic</td>
<td>Yarrow</td>
</tr>
<tr>
<td>Wormwood</td>
<td>Sweet Flag or Calamus</td>
</tr>
<tr>
<td>Tarragon</td>
<td></td>
</tr>
<tr>
<td>Wild Ginger</td>
<td>Roots or root infusions of Indian hemp Calendula</td>
</tr>
<tr>
<td>Goosefoot</td>
<td>Hemp</td>
</tr>
<tr>
<td>Conifers</td>
<td></td>
</tr>
<tr>
<td>Crucifers</td>
<td>Blue cohosh</td>
</tr>
<tr>
<td>Cucurbits</td>
<td>Lady slipper root extract</td>
</tr>
<tr>
<td>Fern</td>
<td>Sweet gale or bog myrtle</td>
</tr>
<tr>
<td>Lupine</td>
<td>Pokeweed</td>
</tr>
<tr>
<td>Nuts</td>
<td>Common knotgrass</td>
</tr>
<tr>
<td>Umbilliferae</td>
<td>Tansy Seeds</td>
</tr>
<tr>
<td>Pyrethrum</td>
<td>Blackberries</td>
</tr>
<tr>
<td>Tobacco</td>
<td>Raspberries</td>
</tr>
<tr>
<td>Beech creosote</td>
<td>Young ash and elder shoots</td>
</tr>
</tbody>
</table>

Areca nut (betel nut), Granatum (pomegranate), Male fern (Aspidium), pepo (pumpkin seed), santonin (levant wormseed) are used as anthelmintics for all animals to expel tapeworm (Karreman, 2004; Hatcher and Wilbert, 1915).

Diatomaceous earth (DE) is the skeletal remains of single-celled algae, or diatoms that formed sedimentary deposits when they died. Diatomaceous earth is comprised predominantly of silicon dioxide. It is a non-synthetic substance and not prohibited for use in organic livestock production (§ 205.105; § 205.237(a); § 205.237(b)). Diatomaceous earth can be added to grain mixes to prevent internal parasite burden in intensively grazed cattle (Karreman, 2004). DE is also used in chicken feed to reduce parasite load from nematodes such as Capillaria aerophila (Bennett et al., 2011). Attempts to use diatomaceous earth to reduce parasite level in goats have not been successful (Bernard et al., 2009). In another study involving sheep, diatomaceous earth mixed with feed, bacillus thuringensis and Clonostachys rosea f. rosea showed efficacy in reducing egg counts for gastrointestinal nematodes (Amhed et al, 2013).

Santonica (Artemisia pauciflora), swamp milkweed (Asclepsia incarnate), brayera or kousso (Brayera anthelmintica), bonduc (Caesalpinia bonduccela), Calumba (Jateorrhiza palmate), Pigella or Maryland pink (Spigella marilandica), and turpentine long-leaved Georgia, swamp or pitch pine (Oleum terebinthiniae) have also shown anthelmintic properties. They are listed in American Materia Medica, Therapeutics and Pharmacognosy with directions for use (Ellingwood, 1919; Karreman, 2004).

Karreman provides a number of references to homeopathic anthelmintic remedies in his book Treating Dairy Cows Naturally: Thoughts and Strategies including Nuzzi, Grainger and Moore, Lust, Levy, Mowry, Dadd, Waterman, Alexander, Burkett, An M.R.C.V.S., Dun, Udall, Winslow and Grosjean (Nuzzi, 1992;
Wormwood (Artemisia absinthium) is known for its ancient use as anthelmintic. The lactones absinthin and anabsinthin are responsible for the anthelmintic activity of wormwood. A. absinthium acts on nicotinic and muscarinic cholinergic receptors (Pepping, 2004).

An in vitro study of susceptibility of Lumbricoides ascaris to a number of plant alcohol extracts disclosed the activities of Acorus calamus (rhizome), Agati gratifola (seeds), Carum copticum (seeds), Cassia tora (seeds), Citrus limonum (seeds), Caesalpinia bonduce (seeds), Curcuma longa (rhizome), Helleborus niger (stem), Mangifera indica (seed kernel) and Zingiber officinale (rhizome) to either paralyze or kill the parasites. From this study, Mangifera indica extracts were used clinically to cure patients (Kaleysa, 1974).

In a study comparing efficacy to control nodular worm (Oesophagostomum spp.) of four medicinal plants fed to pigs with ivermectin treatment sweet flag rhizome (Acorus calamus, 5 grams/kilogram (g/kg)), tansy flowers and leaves (Tanacetum vulgare, 5 g/kg) and pumpkin seeds (Cucurbita pepo, 5 g/kg) reduced worm burden respectively, 98%, 95.8% and 97%, with respect to ivermectin, 96.1% (Magi et al., 2005).

Cassava leaves (Leucaena pallida) added to the diet of goats as a feed additive significantly reduced nematode parasite egg counts and improved weight gain (Merera et al., 2013).

Duddingtonia flagrans is a nematophagous fungus with potential to control trichostrongyles in cattle. Twenty calves, six-month-old, divided in two groups (fungus-treated and control without fungus) were fed on a pasture of Surinam grass known to contain bovine trichostrongyles. Treated animals received sodium alginate mycelial pellets. There was a significant reduction in fecal egg count (56.7%) and infective larvae (L3) in co-procultures (60.5%) in treated animals suggesting that nematophagous fungus might be useful for parasite control (Assis et al., 2012).

**Evaluation Question #12:** Describe any alternative practices that would make the use of the petitioned substance unnecessary (7 U.S.C. § 6518 (m) (6)).

Good husbandry and nutrition are vitally important for good parasite control. The level and quality of feed influences how the animal will cope with parasites, and the level of immunity it will develop against them. Forage crops that support mycorrhizal fungi, and contain high levels of tannins are also good for suppressing parasites (Stockdale, 2008). The use of parasiticides in organic livestock production is meant only as an emergency action to alleviate economic loss and animal suffering (Spoolder, 2007; Charlier et al., 2014).

A number of management practices such as whole-flock treatment of adult ewes around lambing, and treatment of lambs with low parasite contamination as they are moved onto pastures reduces but does not eliminate the use of parasiticides. In addition these practices have been identified as high risk for selecting resistant parasites (Leathwick et al., 2015). Identifying and treating animals that are severely affected by parasites while leaving healthy animals that are coping with the disease untreated and maintaining a reservoir of susceptible parasites has also been effective for reducing the use of parasiticides and suppressing the development of anthelmintic resistance. This is called the FAMACHA system. It provides for a method of identifying diseased sheep using the color of their conjunctiva from deep red in healthy sheep to white in sick sheep as a guide (van Wyk and Bath, 2002). Healthy un-infested animals left untreated in these management systems are still considered organically produced livestock (§205.603(a) (18)). The rule is explicit concerning the treated animal.

In an indoor experiment the development of thiabendazole resistance slowed after exposing smaller proportions of each generation of Haemonchus contortus to treatment with the anthelmintic. Subsequent studies demonstrated that creating a reservoir of unselected parasites, refugia, slows the development of anthelmintic resistance, and emphasizes the risk of treating all animals prior to a shift on to low-contamination pasture. However, higher levels of pasture contamination, resulting from untreated animals, highlight the difficulty in managing both worm control and resistance (Waghorn et al., 2015). Healthy un-infested animals left untreated in these management systems are still considered organically produced livestock (§205.603(a) (18)). The rule is explicit concerning the treated animal.
Grazing management and the use of safe pastures for calves and sheep after weaning is an important component of helminth control in organic farming. It is important to have (1) preventive grazing management such as delayed turn-out, change of pastures between seasons, and the use of more aftermath, (2) diluting grazing management: mixed or alternate grazing with other host species, (3) evasive grazing management like changing the pasture within the season, and (4) supplementary feeding in the spring. Organic farms tend to have a higher diversity of nematodes, since animals are not normally treated with anthelmintic drugs. Helminth diversity has been related to a lower intensity of infection in extensive goat breeding and in meat cattle (Caberet et al., 2002).

Early organic farmers recognized the biological interdependence between animals and plants with the use of a “mixed farming” approach to grazing where (1) animals succeeded one another on the field to avoid species specific transfer of disease, i.e. dairy cattle, then sheep and goats, then beef cattle; (2) only composted animal wastes for fertilizer were used to avoid transfer of known disease agents to the soil and back to their livestock and (3) overcrowding and over grazing were avoided to prevent contact with potentially parasitic worms in various stages of development naturally following bacteria and fungus into specific plants and decomposing material (Sykes, 1949; Ingham, 1999).

Many holistic products are available and effective for worming. Anthelmintic resistance is in part the result of improper use, e.g., the consequence of under dosing, mass therapy and the use of the same class of anthelmintics for prolonged periods of time (Villalba et al., 2014). Resistance to synthetic parasiticides is not a problem, if synthetic parasiticides are not used. Livestock production based on grazing and browsing systems is directly related to the use of plant resources (Alonzo-Diaz, 2014). With proper pasture management, a good diet with plenty of forage for livestock and knowledgeable coaches to provide appropriate strategies for husbandry and treatment healthy animals can be sustainably raised without synthetic parasiticides (Brunetti and Karreman, 2006).

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